



UNIVERSIDAD POLITÉCNICA DE CARTAGENA

**ESCUELA DE INGENIERÍA DE CAMINOS, CANALES Y PUERTOS Y
DE INGENIERÍA DE MINAS**

TRABAJO FIN DE MASTER

**ORGANIC MATTER DYNAMICS AND SOIL
AGGREGATION IN A TECHNOSOL CREATED
WITH METALLIFEROUS MINE RESIDUES, BIOCHAR
AND MARBLE WASTE**

**Autor:
FABIÁN MARCELO MORENO BARRIGA**

MASTER EN CIENCIA Y TECNOLOGÍA DEL AGUA Y DEL TERRENO

Cartagena, Octubre 2016



UNIVERSIDAD POLITÉCNICA DE CARTAGENA

**ESCUELA DE INGENIERÍA DE CAMINOS, CANALES Y PUERTOS Y
DE INGENIERÍA DE MINAS**

TRABAJO FIN DE MASTER

ORGANIC MATTER DYNAMICS AND SOIL AGGREGATION IN A TECHNOSOL CREATED WITH METALLIFEROUS MINE RESIDUES, BIOCHAR AND MARBLE WASTE

**Autor:
FABIÁN MARCELO MORENO BARRIGA**

MASTER EN CIENCIA Y TECNOLOGÍA DEL AGUA Y DEL TERRENO

Cartagena, Octubre 2016

Directores:
José Alberto Acosta Avilés
Ángel Faz Cano
Raúl Zornoza Belmonte

GRATITUDE

Dr. José Alberto Acosta for his support in the preparation of this work

Dr. Raúl Zornoza for all the knowledge shared

Msc. Vicente Diaz for his experimental support and unconditional friendship

INDEX

RESUMEN.....	6
ABSTRACT.....	7
1. INTRODUCTION.....	8
2. OBJETIVES.....	10
2.1 General objctive	10
2.2 Specific objectives.....	10
3. MATERIALS AND METHODS.....	11
3.1 Mine, organic and industrial residues used	11
3.2 Soil incubation	11
3.3 Soil and gas analyses.....	12
3.4 Statisticalanalyses.....	13
4. RESULTS AND DISCUSSION.....	14
4.1 pH and electrical conductivity.....	14
4.2 Carbon and nitrogen dynamics.....	15
4.3 Aggregates stability.....	18
4.4 Microbial biomass and enzymeactivities	19
4.5 Greenhouse gas emissions.....	20
5. CONCLUSIONS	22
6. REFERENCES	24
ANNEX I: TABLES	32
ANNEX II: PHOTOGRAPHS	33

RESUMEN

Los objetivos de este estudio fueron determinar y explicar la influencia del biocarbón y residuos de mármol en dinámica de la materia orgánica y la agregación en una balsa de residuos en el distrito minero de Cartagena-La Unión (SE España). Tres tipos de biocarbón se obtuvieron a partir de estiércol de cerdo, residuos de depuradora municipal y residuos de cosecha hortícola a 500°C durante 2 h. Ocho tipos de tratamientos fueron utilizados, que incluían: un suelo control (CT) sin la aplicación de enmienda, tres tratamientos con diferentes tipos de biocarbón, un tratamiento con los residuos de mármol y tres tratamientos con diferentes biocarbón más residuo de mármol. Se determinaron pH, conductividad eléctrica, agregados estables, carbono orgánico total, carbono inorgánico, carbono recalcitrante, carbono lábil, carbono soluble, nitrógeno total, nitrógeno recalcitrante, nitrógeno lábil, carbono de la biomasa microbiana, β -glucosidasa y actividad arilesterasa, hidrofobicidad, carbohidratos y emisiones de gases de efecto invernadero. Los resultados sugieren que el biocarbón, producido por diversas fuentes es una estrategia eficaz para secuestrar el carbono en el suelo y mejorar la estructura del mismo. Sin embargo, para recuperar de forma efectiva los residuos de mina sería necesario agregar una fuente lábil de materia orgánica con el fin de promover el desarrollo de las poblaciones microbianas.

Palabras clave: enmiendas, biocarbón, residuos de mármol, agregados estables, residuos mineros.

ABSTRACT

The objectives of this study were to determine and explain the influence of biochar and marble waste on soil organic matter dynamics and aggregation in tailings pond at the Mining District of Cartagena-La Union (SE Spain). Three types of biochars were obtained from pig manure, municipal soil waste and crop residue at 500°C during 2h. Eight types of treatments were used, which included: one soil control (CT) without application of amendment, three treatments with different types of biochars, one treatment with marble waste and three treatments with different biochars plus marble waste. pH, electrical conductivity, aggregation stability, total organic carbon, inorganic carbon, recalcitrant carbon, labile carbon, soluble carbon, total nitrogen, recalcitrant nitrogen, labile nitrogen, microbial biomass carbon, β -glucosidase and arylesterase activity, hydrophobicity, carbohydrates and greenhouse gas emissions were determined. The results suggest that biochar, produced by diverse sources, is an effective strategy to sequester carbon in soil and improve soil structure. However, to effectively reclaim mine residues would be necessary to add a labile source of organic matter to promote the development of microbial populations.

Keywords: amendments, biochar, marble waste, aggregation stability, mine waste

1. INTRODUCTION

Metalliferous mine areas constitute degraded ecosystems resulting from mineral extraction for long periods of time, giving rise to large areas of derelict land. Environmental transformations related with mining activities entail changes in the morphology of the area owing to the extraction of minerals and dumping of residues, with reduction of vegetation cover and fauna diversity, and changes in soil quality and structure (Sadhu et al., 2012; Zawadzki et al., 2016). Metalliferous mine residues dumped into the environment have numerous restrictions affecting their development into natural soils, such as extremely low pH, high concentrations of metals and metalloids and extremely low organic matter content (Zanuzzi et al., 2009; Martínez-Pagán et al., 2011). These materials can also provoke acid mine drainage, a dangerous source of water contamination (Barrie and Hallberg, 2005). Therefore, there is a need to develop strategies to reduce the impact of mining residues spread on mine landscapes to guarantee ecosystem reclamation. One effective solution is the creation of Technosols (IUSS, 2014) by addition of different amendments, so that microorganisms and vegetation are able to colonize and grow. For this purpose, the proper selection of amendments to create soil structure is critical, since the most important processes for pedogenesis is the accumulation and cycling of soil organic matter and the formation of stable aggregates (Séré et al., 2010; Huot et al., 2014).

Organic residues are commonly used as amendments because the addition of organic matter can significantly improve soil structure and nutrient status (Senesi et al., 2007; Kayikcioglu, 2012). Biochar production through the pyrolysis of organic residues has recently become an interesting solution for successful soil reclamation due to its high recalcitrant carbon content, which can increase the content of stable organic carbon in soil, contribute to carbon sequestration, and improve soil physical, chemical and biological properties (Glaser et al., 2002; Lehman and Rondon, 2006; Marchetti et al., 2012). Biochar has also emerged as a strategy for reducing greenhouse gas (GHG) emissions. However, the effect of biochar on GHG highly depends on biochar feedstock and pyrolysis conditions, soil type and soil management (Cayuela et al., 2010; Scheer et al., 2011; Taghizadeh-Toosi et al., 2012a,b; Harris et al., 2013). The importance of calcareous materials in supplying carbonates and Ca ions

to neutralize acidity, contributing to buffer capacity and stabilize soil organic matter is well recognized (Stumm and Morgan, 1996; Calvet, 2013; Zornoza et al., 2013). For these reasons, several authors have suggested the use of marble wastes as a source of calcium carbonate for acidic soil remediation (Janjirawuttikul et al., 2011; Tozsin et al., 2014a; Zornoza et al., 2013).

Previous studies have reported the positive effects of Technosols created on mine and industrial residues through the addition of different amendments to ensure true landscape reclamation (Séré et al., 2010; Huot et al., 2014; Wiszniewski, et al 2016). However, to really conclude that the addition of different organic and industrial wastes to mine residues efficiently contributes to soil creation, a proper and thorough monitoring of organic matter stability, GHG emissions, aggregation, and microbial biomass and activity should be developed. Since soil microorganisms are key in the stabilization and degradation of soil organic matter and in the formation of soil stable aggregates (Agegnehu et al., 2016; Zornoza et al., 2016a), the monitoring of microbial biomass and activity, and its interaction with soil physicochemical properties will help gain knowledge about microbial mediated processes in Technosols.

In order to elucidate the main factors controlling organic matter stabilization and degradation and soil aggregates formation in a Technosol derived from mine residues as strategy for reclamation, we performed a short-term incubation experiment with biochars derived from different feedstock and marble waste. Our objective was to assess the effects of the different amendments on the evolution of C and N pools, GHG emissions (CO₂, N₂O and CH₄), aggregate stability, and microbial biomass and activity. We hypothesized that the application of biochar and marble waste to mine soils can contribute to the stabilization of organic matter by formation of stable aggregates and the activation of microbial populations. Biochar type could have a strong influence on aggregate stability and microbial stimulation, while CaCO₃ may have a positive effect on aggregation by release of Ca²⁺ cations.

2. OBJETIVES

2.1 General objective

The objective of this study were to determine and explain the influence of biochar and marble waste on soil organic matter dynamics and aggregation in tailings pond at the Mining District of Cartagena-La Union (SE Spain).

2.2 Specific objectives

The different soils are periodically analyze to monitor the evolution of: pH, electrical conductivity, aggregate stability, total organic carbon, inorganic carbon, recalcitrant carbon, labile carbon, soluble carbon, total nitrogen, recalcitrant nitrogen, labile nitrogen, carbohydrates, microbial biomass carbon, β -glucosidase activity, arylesterase activity, hidrofobicity and greenhouse gas emissions (CO₂, N₂O and CH₄) at 0, 2, 4, 7, 15, 30 and 90 days of incubation.

3. MATERIALS AND METHODS

3.1 Mine, organic and industrial residues used

A tailings pond at the Mining District of Cartagena-La Unión (SE Spain)(37°35'38'' N, 0°53'11" W) was selected to collect the mine residues, which were characterized by high acidity and metal concentrations, low organic carbon and nutrient contents, and sandy loam texture. The climate of the area is semiarid Mediterranean, with mean annual temperature of 18 °C and mean annual rainfall of 275 mm. The mine residue was collected from the top 20 cm of the tailings pond, air-dried for seven days, and sieved <4mm for incubation experiments.

Biochar feedstock was pig manure (PM), cotton (*Gossypiumhirsutum*L.) crop residues (CR) and municipal solid waste (MSW), collected in a pig farm, agricultural field and municipal solid waste treatment plant from the municipality of Cartagena (SE Spain). The feedstock was dried at 60 °C in a forced air lab oven during 72 h and then was ground to pass a 4-mm sieve. The ground particles were then pyrolyzed in a muffle furnace. The temperature was increased at 5 °C min⁻¹ to 500 °C and then maintained for 2 h at this temperature for each feedstock type. Biochar was then ground at < 250 µm for laboratory incubations. The marble waste (MaW; formed by particles of 5-10 µm diam.) was collected from quarries in the Northwest of the Region of Murcia(SE Spain). The main characteristics of all residues used are shown in Table 1 (Annex I).

3.2 Soil incubation

Eight different treatments were applied to the mine residue: unamendment mine residue used as control(CT), PM, CR, MSW, MaW, PM+MaW, CR+MaW and MSW+MaW. Biochars were thoroughly mixed with the mine residue at a dose of 20 g C kg⁻¹ soil, which is the organic C content in the native shrubland soils of the area (Martinez-Martinez et al., 2013).The MaW was added in a rate of 50 g kg⁻¹. This rate was calculated using the method proposed by Sobek et al. (1978), which specifies the amount of CaCO₃ required to neutralize the acidification potential of the soil, according to the percentage of sulfides present in the mine residue, to reach

a final pH of eight, which is the pH value in the native shrubland soils of the area (Martinez-Martinez et al., 2013).

Laboratory incubation (three replicates per treatment) was performed in plastic pots kept in the dark, to a constant humidity of 50% of water holding capacity and temperature of 22 °C under aerobic conditions during 90 days. Soil moisture levels were gravimetrically maintained adding deionized water if needed. Technosols were sampled to monitor the evolution of pH, electrical conductivity (EC), aggregate stability (AS), total organic carbon (TOC), inorganic carbon (IC), recalcitrant carbon (Crec), labile carbon (Clab), soluble carbon (Csol), total nitrogen (Nt), recalcitrant N (Nrec), labile nitrogen (Nlab), carbohydrates, microbial biomass carbon (MBC), β -glucosidase activity, arylesterase activity, hydrophobicity and greenhouse gas (GHG) emissions (CO₂, N₂O and CH₄) at 0, 2, 4, 7, 15, 30 and 90 days of incubation. The first time-point was collected just after rewetting. The CT and MaW treatments were excluded in the analyses for Crec, Nrec, Clab and Nlab, since they had very low values, and experimental error could be higher than samples variability.

3.3 Soil and gas analyses

Soil pH and EC were measured in deionized water (1:2.5 w/v and 1:5 w/v, respectively); AS was determined by the method proposed by (Roldán, 1994); TOC, IC and Nt were determined by an elemental analyzer CNHS-O (EA-1108, Carlo Erba); Crec, Nrec, Clab and Nlab were determined by hydrolysis with H₂SO₄ (Rovira and Vallejo, 2000); the recalcitrance index for carbon (RCI = Crec / Corg) and nitrogen (RNI = Nrec / Nt) was calculated; carbohydrates were extracted in deionized water (1:5 w/v) and measured using the colorimetric method proposed by Brink et al. (1960) by addition of 0.2% anthrone in sulphuric acid; MBC was determined using the fumigation-extraction procedure after extraction with 0.5 M K₂SO₄ (Vance et al., 1987). The non-fumigated fraction was considered as Csol; β -glucosidase activity was determined according to Tabatabai (1982), arylesterase activity was measured according to Zornoza et al. (2009); the hydrophobicity was assessed by the water drop penetration time test (Wessel, 1988).

GHG emissions were measured as the amount of CO₂, N₂O and CH₄ evolved from 2 g of soil in 25 mL glass vials closed with crimp caps after 24 h incubation at 22 °C using a gas chromatograph (AGILENT 6890N).

3.4 Statistical analyses

The fitting of the data to a normal distribution for all properties measured was checked with the Kolmogorov-Smirnov test at $P < 0.05$. Data were transformed using logarithms to assure normal distribution. A one-way repeated measures ANOVA was carried out for all properties independently for each sampling date to assess differences among treatments. Histograms of the residuals from ANOVA were plotted for each variable to confirm that the normality assumption was plausible. When the null hypothesis was rejected, the separation of means amongst levels was made according to Duncan at $P < 0.05$. Relationships among properties were studied using Pearson correlations. Statistical analysis was performed with the software IBM SPSS Statistics 20.

4. RESULTS AND DISCUSSION

4.1 pH and electrical conductivity

All amendments were able to significantly ($P < 0.001$) increase pH owing to their alkaline nature (Fig. 1A). The samples with marble waste showed the highest values (7.76-8.25), confirming that MaW is an effective amendment for pH neutralization, also observed by previous authors (Janjirawuttikul et al., 2011; Tozsin et al., 2014a; Zornoza et al., 2013).

EC only significantly decreased ($P < 0.001$) with regard to CT in the treatments containing MaW, with an average value of 4.21 dS m^{-1} at the end of the incubation (Fig 1.B). The combined addition of biochar with MaW showed significantly higher EC than the treatment that only received MaW, likely due to the presence of soluble salts in the biochar (Zornoza et al., 2016), which can be incorporated into the soil. Increases in EC after addition of biochars derived from different sources have been previously reported owing to the input of soluble salts (Burrell et al., 2016; Molnár et al., 2016). Thus, the decrease in EC was a direct effect of the addition of calcium carbonate, since Ca^{2+} can react with soil sulphates (which can derive from oxidation of pyrite, galena, sphalerite, etc) and form mineral precipitates such as $\text{CaSO}_4 \cdot 2\text{H}_2\text{O}$, which can reduce the amount of soil soluble sulphates (Fernández-Caliani and Barba-Brioso, 2010). The negative correlation between IC and EC supports this fact ($r = -0.89$; $P < 0.001$). The increase in pH can also promote the precipitation of different ions in the soil. In fact, the negative correlation found between pH and CE supports this hypothesis ($r = -0.82$; $P < 0.001$). Although decreases in EC are not essential to contribute to soil formation, they are positive to facilitate the colonization and growth of microbial and plant communities. However, salinity is still high in the Technosol created with the addition of MaW; so, only salt tolerant microorganisms and plant species would satisfactorily grow (USDA, 1999).

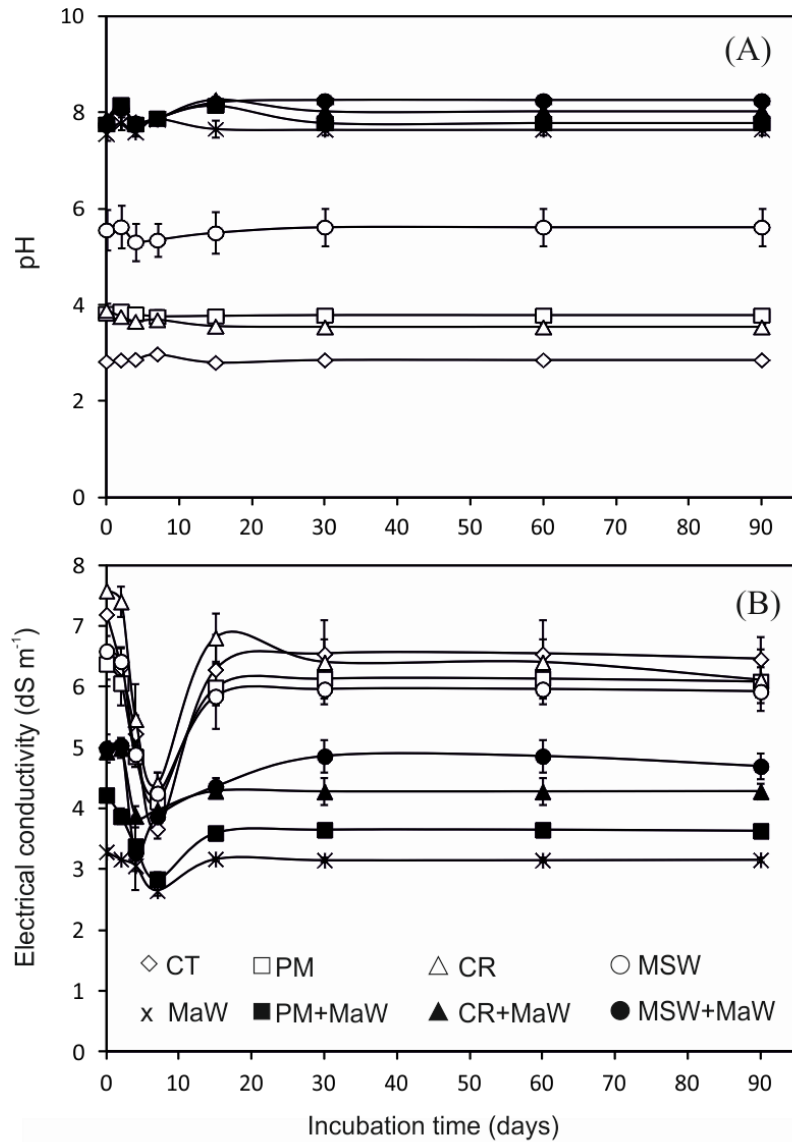


Figure 1. Evolution of pH (A) and electrical conductivity (B) in the unamended mine residue and Technosols created with different amendments during 90 days of incubation. Error bars in each sampling date denotes standard deviation ($n = 3$). CT: control (unamended mine residue); PM: pig manure-derived biochar; CR: crop residue-derived biochar; MSW: municipal solid waste-derived biochar; MaW: marble waste.

4.2 Carbon and nitrogen dynamics

IC was also stable during the entire incubation in those samples receiving MaW, with values $\sim 20 \text{ g kg}^{-1}$ (data not shown), promoting the maintenance of high pH in soils, with significant positive correlation between both properties ($r=0.72$; $P < 0.001$). TOC significantly increased in all treatments receiving biochar with regard to CT and MaW, with an average value

of 18 g kg^{-1} (Fig. 2A). TOC remained stable at this level during the incubation in all samples, which was the organic carbon added with the biochars. Thus, there was no significant loss of organic carbon in any sample receiving biochar, confirming the high stability of the biochars. In fact, RCI was high in all treatments receiving biochar (Fig. 2B), with values > 0.70 . CR-derived biochar, alone or in combination with MaW showed significantly lower RCI (0.70-0.80) than the rest of treatments (0.95-0.97). Clab increased during the first 7 days of incubation in all treatments (Fig. 2C). From that day, Clab slightly decreased and maintained stable during the incubation. Clab was higher in CR-derived biochar, alone or in combination with MaW, than in the other treatments during the incubation. Csol followed the same trend than Clab, with increases up to day 7, to start at that date a decreasing trend (Fig. 2D). However, there were no significant differences among treatments with regard to Csol. Carbohydrates content was below detection limit in all samples. Thus, these results suggest that water-soluble organic compounds provided by biochar are scarce and do not incorporate into the soil. In addition, many soluble organic compounds can be adsorbed in biochar surfaces, and act and cement agents to form aggregates (Liang et al., 2010), and so no increase is detected in soil. Contrarily to our findings, Wang et al. (2016) observed that biochar addition to agricultural soils could increase Csol derived from native SOM, being this increased $< 1\%$ of total biochar C; however, this effect cannot be expected in mine soils with practically null native SOM. Thus, biochar can act as an amendment to increase stable organic carbon in soil, favoring carbon sequestration, with low incorporation of labile and soluble compounds.

Nt followed the same trend than TOC (Fig. 2E), with significant increases ($P < 0.001$) in all treatments receiving biochar with regard to CT and MaW, with an average value of 1.02 g kg^{-1} (Fig. 2F). RNI showed no significant differences among treatments receiving biochar, with values ranging from 0.81 to 0.97. These results suggest that most nitrogen provided by biochar is included in recalcitrant organic compounds, with low availability. RNI showed a negative significant correlation with RCI ($r = -0.71$; $P < 0.001$), suggesting that those soils with higher recalcitrant nitrogen compounds are related to lower recalcitrant carbon compounds. Nlab showed very low values ($< 0.35 \text{ mg kg}^{-1}$) and high variability, with no significant differences

among treatments (Fig. 2G). C/N ratio increased with addition of biochars from 3.8 to ~18.0, with no significant differences among samples receiving biochar (data not shown). This trend confirms the stabilization of organic matter in the Technosol, an essential process for soil creation. No sample showed hydrophobicity at any sampling time (data not shown). Although some biochars can become hydrophobic, those produced at high temperatures like in this experiment lose their hydrophobic compounds (Zornoza et al., 2016b).

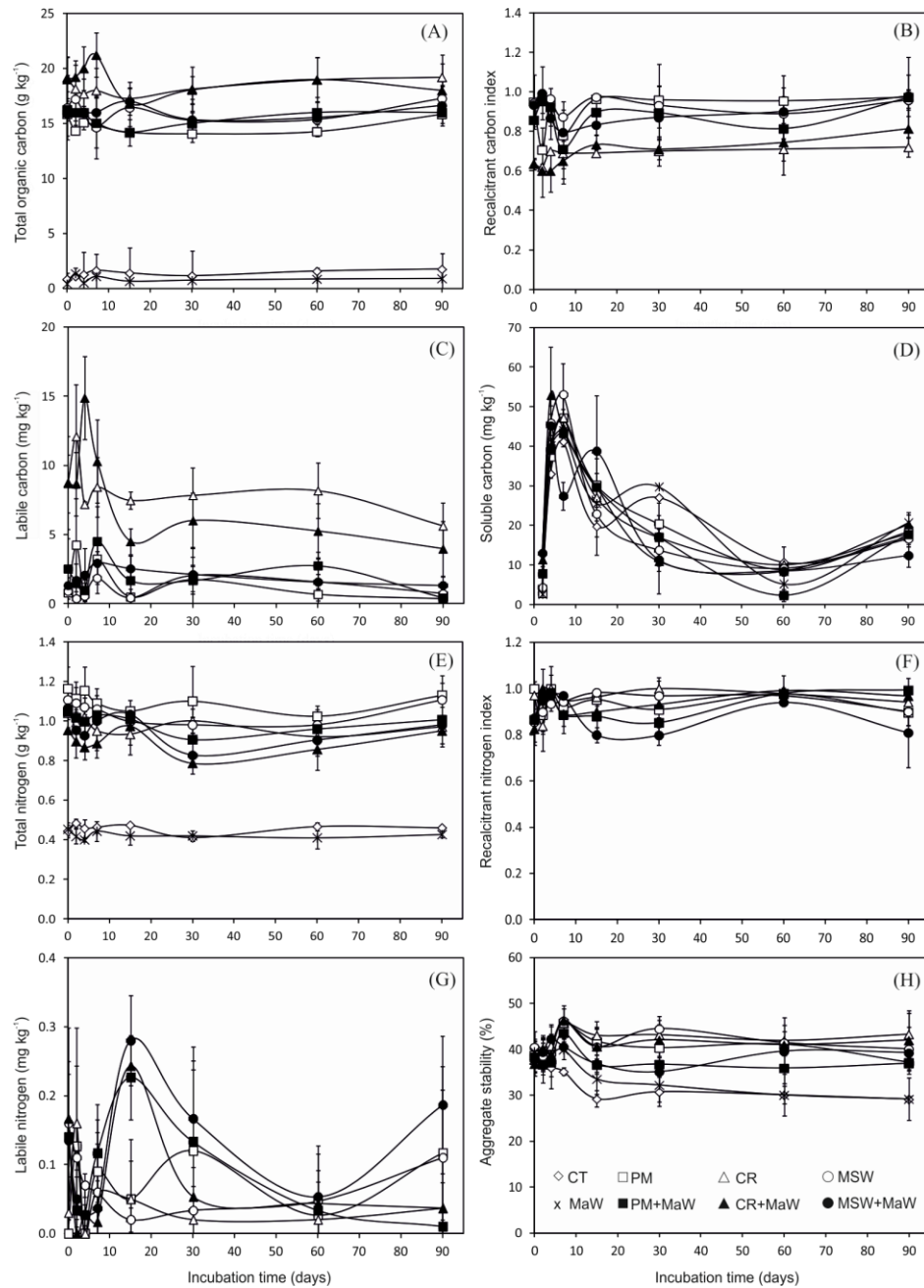


Figure 2. Evolution of total organic carbon (A), recalcitrant carbon index (B), labile carbon (C), soluble carbon (D), total nitrogen (E), recalcitrant nitrogen index (F), labile nitrogen (G) and aggregate stability (H) in the unamended mine residue and Technosols created with different

amendments during 90 days of incubation. Error bars in each sampling date denotes standard deviation ($n = 3$). CT: control (unamended mine residue); PM: pig manure-derived biochar; CR: crop residue-derived biochar; MSW: municipal solid waste-derived biochar; MaW: marble waste.

4.3 Aggregates stability

The different biochars significantly increased ($P < 0.001$) soil aggregate stability with regard to CT by ~40% (Fig. 2H). This increase, not observed with the addition of MaW alone, appeared after 7 days of incubation, and values were practically stable up to the end of the experiment, with no significant differences among the treatments receiving biochar. Biochar acts as a binding agent with minerals, organic matter and microorganisms and thus promoting the formation and stabilization of macro-aggregates (An et al., 2010; Jones et al., 2010). In addition, biochar surface can have labile organic compounds which may act as substrates for microorganisms which, in turn, may build further aggregates through the excretion of mucilage or development of fungal hyphae (Steinbeiss et al., 2009; Liang et al., 2010). It is important to highlight that increases in AS were found after a decrease in C_{lab} and C_{sol} , which may be stabilized by its interaction with mineral particles, or also consumed by microbial populations.

The combined addition of biochar with MaW had no effect on AS. Zanuzzi et al. (2009) concluded in acidic mine residues amended with pig manure and sewage sludge that the addition of calcite was recommended to stabilize soil organic matter and promote the development of soil structure, since the Ca^{2+} bridge with organic compounds favors the formation of stable aggregates. However, since the stabilization of organic matter is needed to increase water-stable macroaggregates (Herath et al., 2013; Zornoza et al., 2016a), it seems that the addition of biochar, with high recalcitrant organic matter, can contribute to increase AS with no extra addition of Ca^{2+} in acidic mine soils. In fact, AS was significantly correlated with TOC ($r=0.63$; $P < 0.001$), N_t ($r=0.52$; $P < 0.001$), C_{rec} ($r=0.54$; $P < 0.001$) and N_{rec} ($r= 0.49$; $P < 0.01$), confirming that the stable organic matter added with biochar was responsible for increments in the AS. Zornoza et al. (2016a) also observed that there was a significant positive relation between soil recalcitrant carbon and aggregate stability in reclaimed mine soils with biochar.

4.4 Microbial biomass and enzyme activities

Biochars significantly increased MBC during the first 7 days with regard to CT (Fig. 3A). However, from this date, there were no significant differences among treatments any longer, with MBC content similar in amended and control samples. This pattern was similar to that observed with Csol (Fig. 2D), suggesting that microorganisms started growing with the presence of easily-available organic substrates (Pérez de Mora et al. 2005), which were rapidly depleted or stabilized after 7 days. Biochar was not able to provide further available substrates for microbial development, and microorganisms size could not increase.

β -glucosidase activity was below detection limit in all samples during the entire incubation. This suggests that microorganisms are not releasing this enzyme related in the hydrolysis of glycosides and cellobiose to obtain monosaccharides, likely due to the lack of free easily available carbohydrates provided by biochar. Some authors previously found that biochar addition can suppress labile organic compounds to enzyme degradation by adsorption on its surfaces (Kasozi et al., 2010; Zimmerman et al., 2011).

Arylesterase activity was more closely related to changes in pH than in organic carbon content. This is confirmed by the significant positive correlation between arylesterase and pH ($r= 0.94$; $P < 0.001$), and the lack of correlations with any organic fraction. Thus, those treatments that received MaW were able to highly increase the activity of this enzyme (Fig. 3B). Zornoza et al. (2009) and Renella et al. (2011) previously reported that arylesterase activity is highly sensitive to shifts in pH, with higher activity in basic soils.

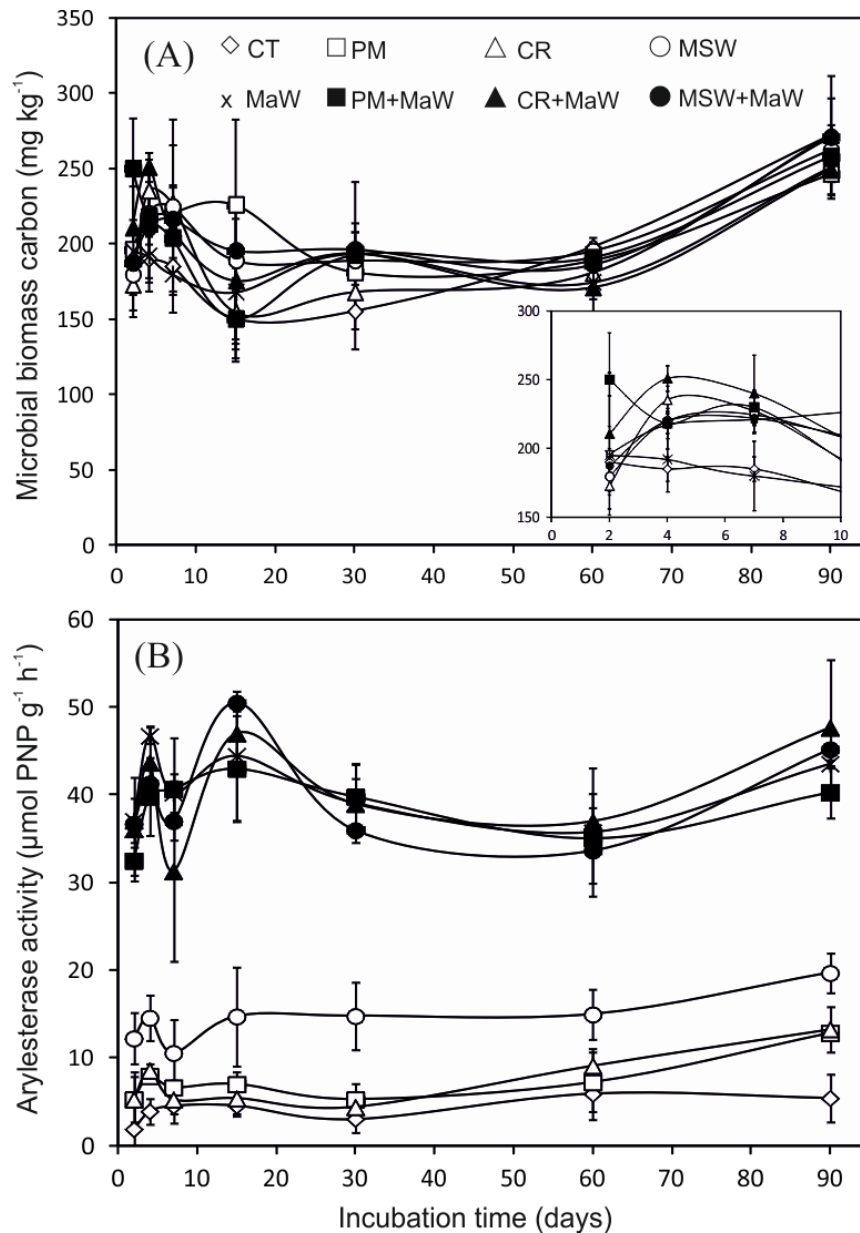


Figure 3. Evolution of microbial biomass carbon (A) and arylesterase activity (B) in the unamended mine residue and Technosols created with different amendments during 90 days of incubation. Error bars in each sampling date denotes standard deviation ($n = 3$). CT: control (unamended mine residue); PM: pig manure-derived biochar; CR: crop residue-derived biochar; MSW: municipal solid waste-derived biochar; MaW: marble waste; PNP: *p*-nitrophenol. Inset in top graph is an enlarged representation of the first 10 days of incubation to visualize shifts in microbial biomass carbon.

4.5 Greenhouse gas emissions

Previous studies have shown biochar amendments to soil have the potential to reduce GHG emissions including CO_2 , ethane (CH_4), and nitrous oxide (N_2O) (Cayuela et al., 2010;

Scheer et al., 2011), this justified the behavior of our amendments at the end of the experimentation.

CO₂ emissions were not significantly affected by any amendment, with values similar to CT (Fig. 4A, B). N₂O emissions increased, as a general pattern, with addition of biochar (Fig. 4C). Increases were only significant for PM+MaW, MSW+MaW and CR during the first 30 days, and for CR and CR+MaW at the end of incubation. When the cumulative N₂O emissions are shown (Fig. 4D), it can be observed that CR, CR+MaW and PM+MaW significantly increased the cumulative N₂O emissions with regard to CT by 16 mg kg⁻¹, 12 mg kg⁻¹ and 10 mg kg⁻¹, respectively after 90 days. Thus, increased N₂O emissions were related to lower RCI. All soils receiving biochar significantly decreased CH₄ emissions from day 30 with regard to CT (Fig. 4E, F), with an average saving of 22 mg kg⁻¹ in 90 days.

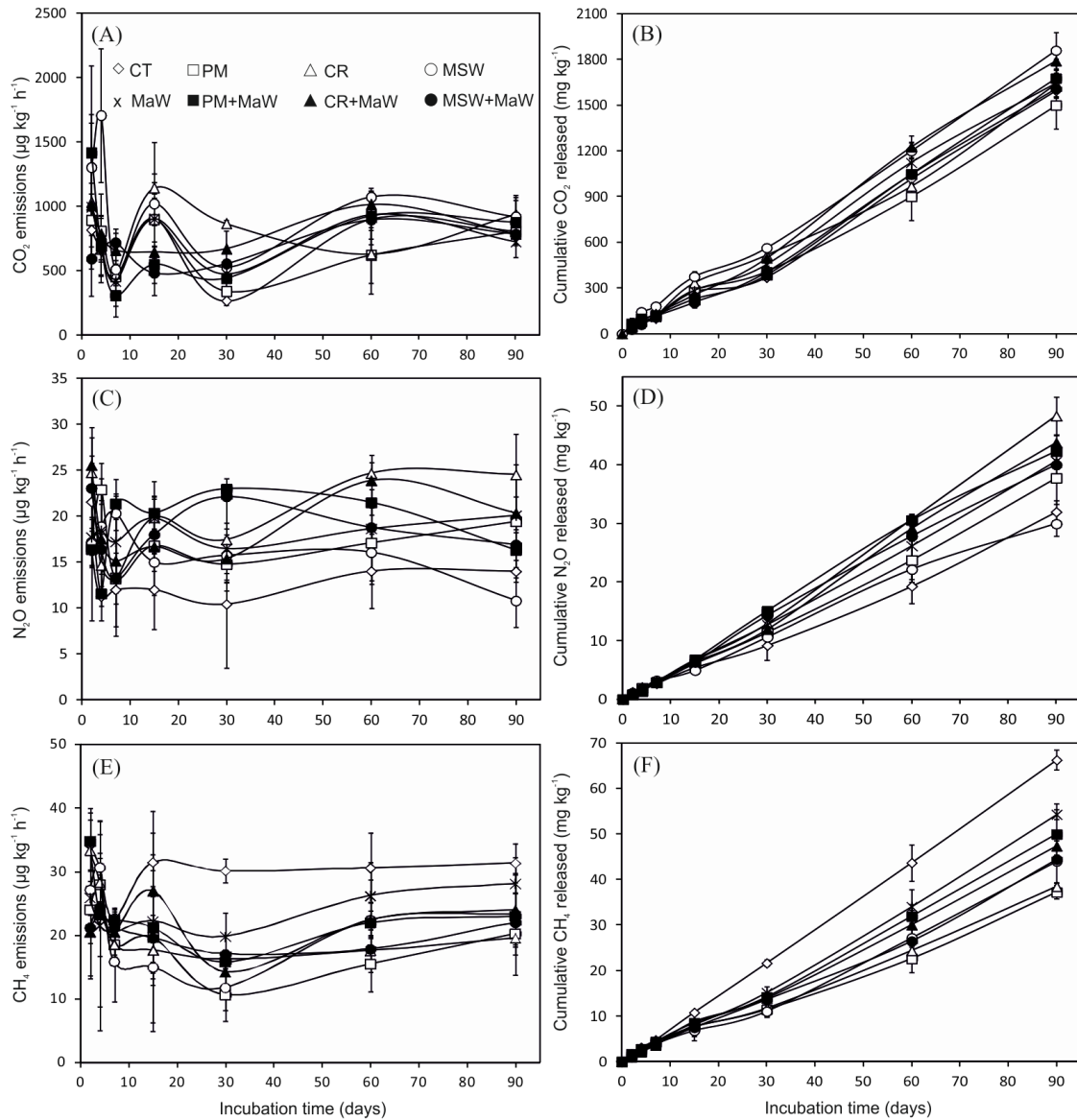


Figure 4. Evolution of CO₂ emissions (A), cumulative CO₂ released (B), N₂O emissions (C), cumulative N₂O released (D), CH₄ emissions (E) and cumulative CH₄ released (F) in the unamended mine residue and Technosols created with different amendments during 90 days of incubation. Error bars in each sampling date denotes standard deviation (n = 3). CT: control (unamended mine residue); PM: pig manure-derived biochar; CR: crop residue-derived biochar; MSW: municipal solid waste-derived biochar; MaW: marble waste.

5. CONCLUSIONS

All the Technosols created with biochar, alone or in combination with marble waste, increased aggregate stability and soil organic carbon and nitrogen, with high recalcitrance indices.

However, the labile and water soluble organic compounds provided with biochars were low to

stimulate microbial biomass and activity. The only effect observed with the addition of marble waste was the increment and maintenance of pH around the target value, the decrease in electrical conductivity, and the improvement of the environment for arylesterase activity. No effect of marble waste was observed in any organic matter fraction neither aggregate stability. All Technosols created were able to reduce CH₄ emissions, with no effect on CO₂emissions. However, N₂O emissions increased with the addition of crop residue-derived biochar alone or together with marble waste, and with the addition of pig manure-derived biochar in combination with marble waste, related to lower recalcitrance index for soil organic carbon. These results suggest that biochar, produced by diverse sources, is an effective strategy to sequester carbon in soil and improve soil structure. However, to effectively reclaim mine residues, it would be necessary to add a labile source of organic matter to promote the development of microbial populations.

Acknowledgements

This work has been funded by Fundación Séneca (Agency of Science and Technology of the Region of Murcia, Spain) by the project 18920/JLI/13.

6. REFERENCES

- Agegnehu G., Bass A., Nelson P., Bird M., Benefits of biochar, compost and biochar–compost for soil quality, maize yield and greenhouse gas emissions in a tropical agricultural soil, *Science of the Total Environment* 543 (2016) 295–306
- Barrie, J., Hallberg, K., 2005. Acid mine drainage remediation options. *Sci. Total Environ.* 338, 3–14.
- An, S., Mentler, A., Mayer, H., Blum, W.E.H., 2010. Soil aggregation, aggregate stability, organic carbon and nitrogen in different soil aggregate fractions under forest and shrub vegetation on the Loess Plateau, China. *Catena* 81, 226–233.
- Brink, R.H., Dubar, P., Linch, D.L., 1960. Measurement of carbohydrates in soil hydrolysates with anthrone. *Soil Science* 89, 157–166.
- Brodowski, S., John, B., Flessa, H., Amelung, W., 2006. Aggregate-occluded black carbon in soil. *Eur. J. Soil Sci.* 57, 539–546.
- Bruun, E.W., Hauggaard-Nielsen, H., Ibrahim, N., Egsgaard, H., Ambus, P., Jensen, P.A., Dam-Johansen, K., 2011. Influence of fast pyrolysis temperature on biochar labile fraction and short-term carbon loss in a loamy soil. *Biomass Bioenergy* 35, 1182–1189.
- Burrell, L.D., Zehtner, F., Rampazzo, N., Wimmer, B., Soja, G., 2016. Long-term effects of biochar on soil physical properties. *Geoderma* 282, 96–102.
- Calvet, R., 2013. *Le sol*, Second ed. Editions France Agricole, Paris.
- Cayuela, M.L., Oenema, O., Kuikman, P.J., Bakker, R.R., Van Groenigen, J.W., 2010. Bioenergy by-products as soil amendments? Implications for carbon sequestration and greenhouse gas emissions. *GCB Bioenergy* 2, 201–213.
- Cross, A., Sohi, S.P., 2011. The priming potential of biochar products in relation to labile carbon contents and soil organic matter status. *Soil Biol. Biochem.* 43, 2127–2134.
- Dold, B., Fontbote, L., 2001. Element cycling and secondary mineralogy in porphyry Cooper tailings as a function of climate, primary mineralogy, and mineral processing. *J. Geochem. Explor.* 74, 3–55.

- Fernández-Caliani, J.C., Barba-Brioso, C., 2010. Metal immobilization in hazardous contaminated soils after marble slurry waste application. A field assessment at the Tharsis mining district (Spain). *J. Hazard. Mater.* 181, 817-826.
- Frouz, J., Elhottová, D., Kuráž, V., Šourková, M., 2006. Effects of soil macrofauna on other soil biota and soil formation in reclaimed and unreclaimed post mining sites: results of a field microcosm experiment. *Appl. Soil Ecol.* 33, 308–320.
- Frouz, J., Prach, K., Pižl, V., Háněl, L., Stary, J., Tajovský, K., Materna, J., Balík, V., Kalčík, J., Řehouňková, K., 2008. Interactions between soil development, vegetation and soil fauna during spontaneous succession in post mining sites. *Eur. J. Soil Biol.* 44,109–121.
- Glaser, B., Lehmann, J., Zech, W., 2002. Ameliorating physical and chemical properties of highly weathered soils in the tropics with charcoal – A review. *Biol. Fertil. Soils* 35, 219–230.
- Griffith, G.S., Bardgett, R.D., 2008. Influence of resource unit distribution and quality on the activity of soil fungi in a particulate medium. *New Phytol.* 148, 143–151.
- Harris, J.A., Bentham, H., Birch, P., 1991. Soil microbial community provides index to progress, and direction of restoration. *Restor. Manag. Notes* 9, 133–135.
- Harris, K., Gaskin, J., Cabrera, M., Miller, W., Das, K., 2013. Characterization and mineralization rates of low temperature peanut hull and pine chip biochars. *Agronomy* 3, 294–312.
- He, Y., Chen, C.R., Xu, Z., Williams, D., Xu, J., 2009. Assessing management impacts on soil organic matter quality in subtropical Australian forests using physical and chemical fractionation as well as ¹³C NMR spectroscopy. *Soil Biology and Biochemistry* 41 (3), 640e650.
- Hilscher, A., Heister, K., Siewert, C., Knicker, H., 2009. Mineralisation and structural changes during the initial phase of microbial degradation of pyrogenic plant residues in soil. *Org. Geochem.* 40, 332e342.

- Herath, H.M.S.K., Camps-Arbestain, M., Hedley, M., 2013. Effect of biochar on soil physical properties in two contrasting soils: an Alfisol and an Andisol. *Geoderma* 209-210, 188–197.
- Holmstrom, H., Ohlander, B., 2001. Layers rich in Fe- and Mn oxyhydroxides formed at the tailings–pond water interface, a possible trap for trace metals in flooded mine tailings. *J. Geochem. Explor.* 74, 189–203.
- Huot, H., Simonnot, M.O., Watteau, F., Marion, P., Yvon, J., De Donato, P., Morel, J.-L., 2014. Early transformation and transfer processes in a technosol developing on iron industry deposits. *Eur. J. Soil Sci.* 65, 470–484.
- IUSS, 2014. World Reference Base for Soil Resources 2014. International soil classification system for naming soils and creating legends for soil maps. *World Soil Resources Reports No. 106*. FAO, Rome.
- Janjirawuttikul, N., Umitsu, M., Tawornpruek, S., 2011. Pedogenesis of acid sulfate soils in the lower central plain of Thailand. *Int. J. Soil Sci.* 6, 77–102.
- Jones, B.E.H., Haynes, R.J., Phillips, I.R., 2010. Effect of amendment of bauxite processing sand with organic materials on its chemical, physical and microbial properties. *J. Environ. Manage.* 91, 2280–2281.
- Kasozi, G.N., Zimmerman, A.R., Nkedi-Kizza, P., Gao, B., 2010. Catechol and humic acid sorption onto a range of laboratory-produced black carbons (biochars). *Environ. Sci. Technol.* 44, 6189–6195.
- Kayikcioglu, H.H., 2012. Short-term effects of irrigation with treated domestic wastewater on microbiological activity of a verticxerofluent soil under Mediterranean conditions. *J. Environ. Manage.* 102, 108e114.
- Kowalenko, C.G., Ihnat, M., 2010. Effects of limestone applications on soil pH and extractable elements in a cauliflower study. *Can. J. Soil Sci.* 90 (4), 655–665.
- Kuzyakov, Y., Bogomolova, I., Glaser, B., 2014. Biochar stability in soil: decomposition during eight years and transformation as assessed by compound-specific C-14 analysis. *Soil Biol. Biochem.* 70, 229e236.

- Kuzyakov, Y., Subbotina, I., Chen, H.Q., Bogomolova, I., Xu, X.L., 2009. Black carbon decomposition and incorporation into soil microbial biomass estimated by C-14 labeling. *Soil Biol. Biochem.* 41, 210e219.
- Lehmann, J., Rondon, M., 2006. Biochar soil management on highly weathered soils in the humid tropics. In: Ball, A.S., Fernandes, E., Herren, H., Husson, O., Laing, M., Palm, C., Pretty, J., Sanchez, P., Sanginga, N., Thies, J. (Eds.), *Biological Approaches to Sustainable Soil Systems*. CRC Press, Boca Raton, FL, pp. 517–530.
- Lucas, S.T., D'Angelo, E.M., Williams, M.A., 2014. Improving soil structure by promoting fungal abundance with organic soil amendments. *Appl. Soil Ecol.* 75, 13–23.
- Liang, B., Lehmann, J., Sohi, S.P., Thies, J.E., O'Neill, B., Trujillo, L., Gaunt, J., Solomon, D., Grossman, J., Neves, E.G., Luizão, F.J., 2010. Black carbon affects the cycling of non-black carbon in soil. *Org. Geochem.* 41, 206 Geoc
- Luo, Y., Durenkamp, M., De Nobili, M., Lin, Q., Brookes, P.C., 2011. Short term soil priming effects and the mineralisation of biochar following its incorporation to soils of different pH. *Soil Biol. Biochem.* 43, 2304e2314.
- Lehmann, S. Joseph, An introduction, in: J. Lehmann, S. Joseph (Eds.), *Biochar for Environmental Management*, Earthscan, London, 2009, pp. 1e9.
- Marchetti, R., Castelli, F., Orsi, A., Sghedoni, L., Bochicchio, D. 2012. Biochar from swine manure solids: influence on carbon sequestration and Olsen phosphorus and mineral nitrogen dynamics in soil with and without digestate incorporation. *Ital. J. Agron.* 7, 189-195.
- Martínez-Martínez, S., Acosta, J.A., Faz, A., Carmona, D.M., Zornoza, R., Cerda, C., 2013. Assessment of the lead and zinc contents in natural soils and tailing ponds from the Cartagena-La Unión mining district, SE Spain. *J. Geochem. Explor.* 124, 166-175.
- Martínez-Pagán, P., Faz, A., Acosta, J.A., Carmona, D.M., Martínez-Martínez, S., 2011. A multidisciplinary study for mining landscape reclamation: A study case on two tailing ponds in the Region of Murcia (SE Spain). *Phys. Chem. Earth* 36, 1331-1344.

- Molnár, M., Vaszita, E., Farkas, E., Ujaczki, E., Fekete-Kertész, I., Tolner, M., Klebercz, O., Kirckeszner, C., Gruiz, K., Uzinger, N., Feigl, V., 2016. Acidic sandy soil improvement with biochar - A microcosm study. *Science of the Total Environment* 563-564, 855-865.
- Pérez de Mora, A., Ortega-Calvo, J.J., Cabrera, F., Madejón, E., 2005. Changes in enzyme activities and microbial biomass after “in situ” remediation of a heavy metal-contaminated soil. *Appl. Soil Ecol.* 28, 125–137.
- Neff, J.C., Townsend, A.R., Gleixner, G., Lehman, S.J., Turnbull, J., Bowman, W.D., 2002. Variable effects of nitrogen additions on the stability and turnover of soil carbon. *Nature* 419, 915e917.
- Novak, J.M., Busscher, W.J., Watts, D.W., Laird, D.A., Ahmedna, M.A., Niandou, M.A.S., 2010. Short-term CO₂ mineralization after additions of biochar and switchgrass to a Typic Kandudult. *Geoderma* 154, 281e288.
- Renella, G., Zornoza, L., Landi, M., Mench, M., Nannipieri, P., 2011. Ayrlesterase activity in trace element contaminated soils. *European Journal of Soil Science* 62, 590-597.
- Roldán, A., García-Orenes, F., Lax, A., 1994. An incubation experiment to determine factors involving aggregation changes in an arid soil receiving urban refuse. *Soil Biol. Biochem.* 26, 1699–1707.
- Rovira, P., Vallejo, V.R., 2000. Examination of thermal and acid hydrolysis procedures in characterization of soil organic matter. *Commun. Soil Sci. Plant* 31, 81–100.
- Sadhu, K., Adhikari, K., Gangopadhyay, A., 2012. Effect of mine spoil on native soil of Lower Gondwana coal fields: Raniganj coal mines areas, India. *Int. J. Environ. Sci.* 2, 1675–1687. doi:<http://dx.doi.org/10.6088/ijes.00202030052>.
- Santos, F., Torn, M.S., Bird, J.A., 2012. Biological degradation of pyrogenic organic matter in temperate forest soils. *Soil Biol. Biochem.* 51, 115e124.
- Scheer, C., Grace, P., Rowlings, D., Kimber, S., Van Zwieten, L., 2011. Effect of biochar amendment on the soil-atmosphere exchange of greenhouse gases from an intensive subtropical pasture in northern New South Wales, Australia. *Plant Soil* 345, 47–58.

- Séré, G., Schwartz, C., Ouvrard, S., Renat, J.-C., Watteau, F., Villemin, G., Morel, J.-L., 2010. Early pedogenic evolution of constructed Technosols. *J. Soils Sediments* 10, 1246–1254.
- Senesi, N., Plaza, C., Brunetty, G., Polo, A., 2007. A comparative survey of recent results on humic-like fractions in organic amendments and effects on native soil humic substances. *Soil Biol. Biochem.* 39, 1244–1262.
- Singh, N., Abiven, S., Maestrini, B., Bird, J.A., Torn, M.S., Schmidt, M.W.I., 2014. Transformation and stabilization of pyrogenic organic matter in a temperate forest field experiment. *Glob. Change Biol.* 20, 1629e1642.
- Sobek, A.A., Schuller, W.A., Freeman, J.R., Smith, R.M., 1978. Field and laboratory methods applicable to overburdens and mine soils. EPA-600/2-78-054.
- Sparling G P. 1992. Ratio of microbial biomass carbon to soil organic carbon as a sensitive indicator of changes in soil organic matter. *Aust J Soil Res.* 30: 195–207.
- Steinbeiss, S., Gleixner, G., Antonietti, M., 2009. Effect of biochar amendment on soil carbon balance and soil microbial activity. *Soil Biol. Biochem.* 41, 1301-1310.
- Staddon, P.L., Thompson, K., Jakobsen, I., Grime, J.P., Askew, A.P., Fitter, A., 2003. Mycorrhizal fungal abundance is affected by long-term climatic manipulations in the field. *Global Change Biology* 9 (2), 186e194.
- Stumm, W., Morgan, J.J., 1996. *Aquatic Chemistry*. Wiley.
- Tozsin, G., Arol, A.I., Oztas, T., Kalkan, E., 2014. Using marble wastes as a soil amendment for acidic soil neutralization. *J. Environ. Manage.* 133, 374–377.
- Tabatabai, M.A., 1982. Soil enzymes. In: Page, A.L., Miller, R.H., Keeney, D.R. (Eds.), *Methods of Soil Analysis, Part 2*, second ed. ASA and SSSA, Madison, pp. 501–538.
- Taghizadeh-Toosi, A., Clough, T., Sherlock, R., Condron, L., 2012a. Biochar adsorbed ammonia is bioavailable. *Plant Soil* 350, 57–69.
- Taghizadeh-Toosi, A., Clough, T., Sherlock, R., Condron, L., 2012b. A wood based low temperature biochar captures NH₃-N generated from ruminant urine-N, retaining its bioavailability. *Plant Soil* 1–12.

- Tozsin, G., OztasT., Arol, A., Kalkan, E., 2014a, Changes in the chemical composition of an acidic soil treated with marble quarry and marble cutting wastes. *Chemosphere* 138 (2015) 664–667.
- Tozsin, G., Arol, A.I., Oztas, T., Kalkan, E., 2014b. Using marble wastes as a soil amendment for acidic soil neutralization. *J. Environ. Manage.* 133, 374–377.
- USDA. 1999. Soil quality Test Kit Guide. United States Department of Agriculture (ed.) Washington. 82pp.
- Vance, E.D., Brookes, P.C. and Jenkinson, D.S. 1987. An extraction method for measuring soil microbial biomass C. *Soil Biol. Biochem.* 19: 703--707.
- Verheijen, F.G.A., Jeffery, S., Bastos, A.C., van der Velde, M., Diafas, I., 2009. Biochar application to soils – A critical scientific review of effects on soil properties, processes and functions. EUR 24099 EN, Office for the Official Publications of the European Communities, Luxembourg.
- Wang, D., Griffin, D.E., Parikh, S.J., Scow, K.M., 2016. Impact of biochar amendment on soil water soluble carbon in the context of extreme hydrological events. *Chemosphere* 160, 287-292.
- Wang, Y., Zhang, J.H., Zhang, Z.H., 2015. Influences of intensive tillage on water-stable aggregate distribution on a steep hillslope. *Soil and Tillage Research* 151, 82–92.
- Wardle, D.A., Nilsson, M.C., Zackrisson, O., 2008. Fire-derived charcoal causes loss of forest humus. *Science* 320, 629e629.
- Wessel, A.T., 1988. On using the effective contact angle and the water drop penetration time for classification of water repellency in dune soils. *Earth Surf. Proc. Land* 13, 555e562.
- Wiszniewsk, A., Hanus, E., Muszynsk, E., Ciarkowski, K., 2016, Natural Organic Amendments for Improved Phytoremediation of Polluted Soils: A Review of Recent Progress, *Pedosphere* 26(1): 1–12, 2016
- Zanuzzi, A., Arocena, J.M., van Mourik, J.M., Faz, A., 2009. Amendments with organic and industrial wastes stimulate soil formation in mine tailings as revealed by micromorphology. *Geoderma* 154, 69-75.

- Zawadzki, J., Przezdziecki, K., Miatkowski, Z., 2016. Determining the area of influence of depression cone in the vicinity of lignite mine by means of triangle method and LANDSAT TM/ETM+ satellite images. *J. Environ. Manage.* 166, 605–614. doi:<http://dx.doi.org/10.1016/j.jenvman.2015.11.010>.
- Zimmerman, A.R., 2010. Abiotic and microbial oxidation of laboratory-produced black carbon (biochar). *Environ. Sci. Technol.* 44, 1295e1301.
- Zimmerman, A.R., Gao, B., Ahn, M.Y., 2011. Positive and negative carbon mineralization priming effects among a variety of biochar-amended soils. *Soil Biol. Biochem.* 43, 1169-1179.
- Zornoza, R., Faz, A., Carmona, D.M., Acosta, J.A., Martínez-Martínez, S., de Vreng, A., 2013. Carbon mineralization, microbial activity and metal dynamics in tailing ponds amended with pig slurry and marble waste. *Chemosphere* 90, 2606-2613.
- Zornoza, R., Acosta J.A., Faz A., E. Bååthb., 2016a. Microbial growth and community structure in acid mine soils after addition of different amendments for soil reclamation, *Geoderma* 272 (2016) 64–72.
- Zornoza, R., Landi, L., Nannipieri, P., Renella, G., 2009. A protocol for the assay of aryl esterase activity in soil. *Soil Biol. Biochem.* 41, 659–662.
- Zornoza, R., Moreno-Barriga, F., Acosta, J.A., Muñoz, M.A., Faz, A., 2016b. Stability, nutrient availability and hydrophobicity of biochars derived from manure, crop residues, and municipal solid waste for their use as soil amendments. *Chemosphere* 144, 122e130.

ANNEX I: TABLES

Table 1. Main characteristics of the different residues and biochars used for the Technosol creation.

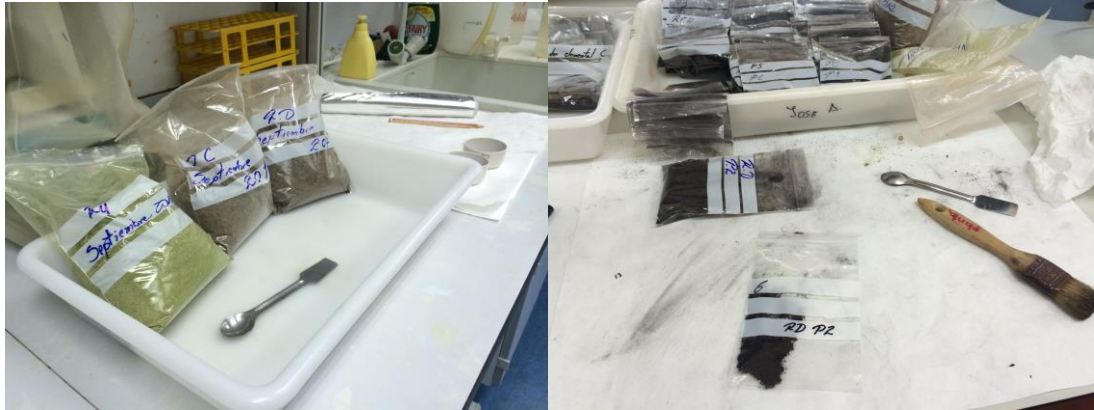
Residue ^a	pH	EC ^b	OC	N	Cu	Zn	Pb	Cd	Ca	Mg	Na	K	CaCO ₃
		dS m ⁻¹	g kg ⁻¹	g kg ⁻¹	mg kg ⁻¹	mg kg ⁻¹	mg kg ⁻¹	mg kg ⁻¹	mg kg ⁻¹	mg kg ⁻¹	mg kg ⁻¹	mg kg ⁻¹	%
Mine residue	2.82	7.20				1030	6636	0.99	-	-	-	-	-
MaW	8.00	2.20	-	-	0.4	260	1.4	0.90	2190	347.00	69.02	59.02	98
PM	10.52	3.99	405	27	100.2	113	3.2	0.35	109	21.27	7.55	24.74	-
CR	10.77	15.89	480	23	16.7	82	10.3	0.10	88	21.01	2.60	68.66	-
MSW	9.78	8.41	288	13	130.6	412	132.1	0.79	142	11.96	13.30	17.98	-

^a MaW: marble waste; PM: pig manure-derived biochar; CR: crop residue-derived biochar; MSW: municipal solid waste-derived biochar

^b EC: electrical conductivity; OC: organic carbon; Nt: total nitrogen

ANNEX II: PHOTOGRAPHS

Source material used and obtained biochars



pH analysis



Stable aggregates analysis



Microbial Biomass Activity analysis



Green House Gases analysis



Mine residues used

