

# Growth and phenological stages of ‘Búlida’ apricot trees in south-east Spain.

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**Short running title:** Phenological stages of apricot trees

## Summary

The different phenological stages of mature apricot trees (*Prunus armeniaca* L., cv. Búlida), drip irrigated and grown under typical Mediterranean conditions are described using the traditional nomenclature of Baggiolini and according to the BBCH General Scale. The predominant dates of each stage are indicated in days and as the number of accumulated growing degree hours (GDH). When the annual pattern of root, shoot and fruit growth was studied, alternating root and shoot growth was evident, and also a relative separation between the main periods of shoot and fruit growth, which is an advantageous characteristic when applying deficit irrigation. This study will improve apricot culture by expressing the timing of most agricultural operations on a standardised scale.

**Key words:** Apricot, growth pattern, *Prunus armeniaca*, phenological stages

**RESUMÉ-** Stades phénologiques et croissance végétative de l’abricotier.

Les stades phénologiques de l'abricotier (*Prunus armeniaca* L., cv. Búlida) irrigué en goutte à goutte et cultivé dans des conditions méditerranéennes ont été étudiés d'après les stades de Bagliolini et le BBCH General Scale. Les dates prédominantes de chacun des stades-repères sont indiquées en degré-heure de croissance (growing degree hours, GDH). Si l'on considère la croissance annuelle des racines, des rameaux et des fruits, on constate une croissance alternative des racines et des rameaux, et aussi une nette séparation entre la période de croissance principale des rameaux et des fruits. Cette étude va améliorer la culture de l'abricotier avec l'utilisation de l'échelle standard des techniques agronomiques.

Abricotier / croissance / *Prunus armeniaca* / stades phénologiques

## 1. INTRODUCTION

In agriculture, a knowledge of the growth factors (water, fertilizers, etc.) involved in a particular crop and the annual pattern of crop growth is essential. A study of the phenological stages of growth in trees is very important not only to predict when hormones should be applied and the possible appearance of disease [3] but also to know the sensitivity to water deficit at each stage of development [46], and to adjust the fertilization programmes to the nutritional needs of the plants (especially nitrogen), thus contributing to a more sustainable agriculture.

Regulated deficit irrigation is increasingly proposed for the optimisation of water use in fruit trees in areas with scarce water resources [12, 20, 21, 22, 34]. This strategy is based on the profound knowledge of phenological plant processes, and its success depends on a clear definition of what is the critical phenological

period for water deficit, when yield and/or fruit quality may be adversely affected [40]. There is abundant information concerning the critical periods, when irrigation should be applied to fulfil total requirements; in the case of fruits trees these period are related with certain stages of fruit growth [12, 24, 25, 26, 43, 46] and with the postharvest period [40, 28]. Also, it is important to know the separation in time between vegetative and fruit growth for specific trees when applying regulated deficit irrigation, because water deficit will only affect one of these processes [10]. In this sense, Chalmers et al. [9] indicated that irrigation management can be a powerful tool to manipulate plant growth for greater fruitfulness and less vegetative growth.

There have been some interesting studies on the floral biology of apricot cultivars under Mediterranean conditions, dealing with cold and heat requirements [8, 29], and physiological processes [17, 39, 40].

Research into the relationship between root and shoot growth of trees has provided conflicting results because data concerning species, sizes and climates, etc. cannot be extrapolated [23]. The use of a standard scale to describe the growth stages of crops is a common solution, but a universal scale, which uses a consistent set of numeric codes adaptable to all crops, is needed. Such is the decimal scale, developed jointly by BASF, Bayer, Ciba-Geigy and Hoechst, named the BBCH General Scale, which identifies different developmental stages by a two-digit code [27].

For these reasons, the aim of this paper was to define the phenological stages of Búlida apricot trees, describing the different growth stages using the traditional nomenclature [5] as well as the BBCH code [27, 32]. This information

will improve the cultivation of this crop in Murcia Region (which produces 58 % of the total Spanish production) by expressing the timing of most agricultural operations on a standardised scale.

## **2. MATERIALS AND METHODS**

### **2.1. Plant material and experimental site**

The experiment was conducted during two growing seasons (1997 and 1998) in a 2 ha plot of a commercial orchard, located in Mula valley, Murcia (SE Spain), with a loam texture soil, highly calcareous, 7.8 pH, and with low organic matter and cationic exchange capacity values.

During the experimental period the climate was typically Mediterranean, with mild winter and dry summer (Table I). The mean daily evaporation rate from a U.S. Weather Bureau class A pan (on bare soil and located at a weather station in the orchard) ranged from 1.4 mm day<sup>-1</sup> in December-January to 7.5 mm day<sup>-1</sup> in July. The annual evaporation for the experimental period averaged 1470 mm, with only minor year-to-year deviations from these values. Annual rainfall averaged 320 mm. The rainy period was as usual in this area, occurring during spring and autumn (Table I).

The plant material consisted of twelve-year-old apricot trees (*Prunus armeniaca* L., cv. Búlida, on Real Fino apricot rootstock), spaced 8 x 8 m, with an average height of 4.5 m, ground cover of 52 % and leaf area index (LAI) of 1.69. The shape of the trees resulted from an open-centre tree training-pruning system. Annually, trees were manually pruned in spring and at the beginning of the rest period (November) in order to eliminate suckers from branches and old wood,

respectively. Trees were drip irrigated using one drip irrigation line for each row, with seven emitters per tree, each with a flow rate of  $4 \text{ l h}^{-1}$ . The experimental design was a randomised block, with four blocks. Each block consisted of two rows of seven trees. The central five trees of the second row were used for experimental measurements (control trees), and the others served as guard trees.

Irrigation amounts were scheduled weekly based on crop coefficients [2], reference crop water use ( $E_{To}$ ), as determined from data collected the previous week in a class A pan, and the estimated application efficiency (95 %). The water amounts applied for the control treatment averaged  $7154 \text{ m}^3 \text{ ha}^{-1} \text{ year}^{-1}$ , maintaining the soil close to field capacity values in the main root zone, and a predawn leaf water potential of around  $-0.45 \text{ MPa}$  (data not shown),

Trees were fertilised with  $164 \text{ kg N}$ ,  $60 \text{ kg P}_2\text{O}_5$  and  $118 \text{ kg K}_2\text{O}$ , per ha per year. A routine pesticide programme was maintained. No weeds were allowed to develop within the orchard, resulting in a clean orchard floor for the duration of the experiment.

Chill requirements were 460 hours, calculated according to Sánchez-Capuchino [42] as the number of hours below  $7 \text{ }^\circ\text{C}$ , and 600 chill units, according to the method of Erez and Couvillon [15]. Harvest was carried out following local commercial criteria on several dates during one month, the first pick taking place in mid-May (15th May, 1997 and 20th May, 1998). The average yield for the studied period was  $256 \text{ kg tree}^{-1}$  ( $5300 \text{ fruits tree}^{-1}$ ), which is more than three times the apricot yield of commercial plantations in the area, which points to the good degree of orchard management, mainly due to the drip irrigation scheduling applied.

## 2.2. Measurements

Following a fixed plan, phenological and growth measurements were made on the same dates in 1997 and 1998. The different phenological stages of apricot growth were defined according to the BBCH General Scale [32], while reproductive phenological stages were described according to Baggiolini [5]. To record the phenological stages, as well as the duration of every stage, four healthy trees were selected at random from control trees of each block. From each tree, four two-year-old branches (1 m long and 1.5 cm<sup>2</sup> in diameter, each containing 150-200 bud flowers) for each compass direction, were tagged. Twice a week, from the end of January (at stage A, dormant buds) to leaf fall in December the different phenological stages were recorded separately. From fruit-set to harvest, counting was only carried out once a week.

The shoot length of four tagged shoots per tree, one from each compass direction, was measured on one tree per block every 14 days, while the trunk diameter in all control trees (five per block) was measured every two months, 30 cm above the soil line. The diameter of 10 tagged fruits per tree was measured weekly in all control trees per block using an electronic digital calliper.

The minirhizotron method was used for root growth measurement, as described by Abrisqueta et al. [1]. One minirhizotron tube per tree (one per block) was placed in the wetted area and root length per unit of soil volume was obtained according to Upchurch and Ritchie [47].

The influence of temperature on the duration of each phenological stage was studied using the growing degree hour (GDH) model [37], calculating the

GDH's by subtracting 6°C (base temperature according to Tabuenca and Herrero [44]) from each hourly temperature.

The physiological fall of buds, flowers, fruits and leaves was evaluated using catching nets of 1/8 of tree canopy (3.5 m radius) installed under one tree per block. Weekly, the different organs were collected, cleaned and separately weighed (fresh). The number of each organ in a sub sample of about 20 g was counted and the dry weight was determined, calculating the total number per net using the fresh/dry weight ratio.

### **3. RESULTS**

#### **3.1. Phenological stages of development**

The phenological growth stages of Búlida apricot trees, according to the BBCH and Baggiolini codes, are shown in Table II and illustrated in Photograph 1. The growing degree hour (GDH) accumulated at the beginning of each stage is also shown in Table II. The description of the main stages (first digit) and some of the secondary stages (second digit), both scaled 0 to 9, is as follows:

##### **Stage 0. Bud development.**

**00**: completely closed bud, corresponding to stage A of the Baggiolini code. With increased temperatures, vegetative bud swelling begins (**01**), starting when 1035 °C GDH has been accumulated. **07**: beginning of bud burst, and **09**: buds open.

##### **Stage 1. Leaf development.**

This covers the time from which the first leaves emerge (**10**), expanding leaves (**11-15**), to **19** (fully expanded leaves), which occurred during mid March,

starting with 5150 °C GDH.

### **Stage 3. Shoot development**

**31:** beginning of the first shoot growth, starting at the beginning of March, with 2334 °C GDH accumulated. This occurred simultaneously with the leaf development (stage 1). **32, 35, 37:** shoots about 20, 50, and 70 % of the final length, respectively. **39:** second shoot growth, initiated at the end of May.

### **Stage 5. Flower emergence**

**51:** flower bud swelling, corresponding to stage B of the Baggiolini code, which occurred during the second half of February, with 1827 °C GDH accumulated. **55:** calyx perceptible, corresponds to stage C of the Baggiolini code, which lasted one week and started with 1447 °C GDH. **58:** flower petals perceptible, corresponding to Baggiolini stage D; this stage was very ephemeral (2 days) and started with 3274 °C GDH, and **59:** anthers perceptible, corresponding to stage E of the Baggiolini code.

### **Stage 6. Flowering.**

**61:** beginning of flowering (10 % of flowers open). **65:** full bloom, corresponding to Baggiolini stage F (50 % of flowers open), and occurring during the first week of March, with 3784 °C GDH. **67:** flowers fading, corresponding to stage G of the Baggiolini code, and **69:** end of flowering, which occurred during mid March, with 4725 °C GDH.

### **Stage 7. Fruit development.**

**71:** fruit set, corresponding to stage H of the Baggiolini code; this stage lasted 13 days and occurred with 4725 °C GDH. **72:** green young fruits, corresponding to Baggiolini stage I. **73:** physiological fruit drop, starting at the



beginning of April, with 9269 °C GDH. **75** and **79**: fruits about 50 and 90 % of the final size, respectively.

#### **Stage 8. Fruit maturity.**

**81**: colour-break; this occurred at the end of April, when 15124 °C GDH had been accumulated. **89**: fruit ripe for consumption.

#### **Stage 9. Senescence.**

**93**: beginning of senescence, starting with 86139 °C GDH accumulated. **95**: leaves starting to fall. **97**: total leaf abscission (winter dormancy period), occurring in mid December, with 93367 °C GDH accumulated.

The accumulation of GDH at full bloom (65 BBCH code, Table II) was 3584 and 3983 °C GDH in 1997 and 1998, respectively, which resulted in a difference of three days between the respective full bloom (5 and 8 March). It must be pointed out that in 1996 full bloom occurred on 20 March (data not shown), which implied a delay of 15-12 days with respect to 1997 and 1998, respectively, but GDH accumulation was 3935 °C GDH, with a minor deviation from the mean value recorded in Table II ( $3834 \pm 218$ ). This fact was mainly due to the cooler February temperatures registered in 1996 with a GDH accumulation during the second fortnight of 1405 °C GDH, unlike the figures seen for 1997 (2523 °C GDH) and 1998 (2145 °C GDH).

The first pick of the harvest does not refer to a BBCH code, because it followed local commercial criteria. However, this would correspond to something between 81 and 89 BBCH code (Table II). GDH accumulation for this cultural procedure occurred at an average of 20859 °C GDH for 1997 and 1998. In 1996

apricot were first picked on 24 May, when 20439 °C GDH were accumulated, which represented a delay of 12 days with respect to 1997 (with 19197 °C GDH accumulated).

The evolution of the reproductive stages of Búlida apricot, according to the Baggiolini code is shown in *figure 1*. It is clear that the progress from stage B (flower bud swelling) to stage F (full bloom) was very rapid, lasting less than 20 days, all stages occurring simultaneously during 8 days. From full bloom (stage F) to young fruit (stage I) lasted 20 days, with less overlapping of these stages.

### **3.2. Pattern of root, shoot and fruit growth**

The annual pattern of root, shoot and fruit growth of apricot trees is shown in *figure 2*. Root growth was active all year, with some fluctuations. In February, immediately before full bloom, there was an important increase in root growth, while maximum root growth activity occurred in April. Minimum values were reached during summer and similar values were observed in autumn-winter.

Shoots had two periods of active growth, both of a short duration (*figure 2*). The first, which was the more intense, occurred after flowering (mid March) and the second after harvest (mid June), affecting only long-type shoots. Both periods coincided with the minimum values of root growth (*figure 2*).

Fruits showed two periods of active growth (*figure 2*), the first at the end of March and the second at the end of May, separated by the lag phase of slower growth and dominated by lignification of the endocarp. The first period occurred after the first shoot growth period (*figure 2*) and coincided with maximum root growth (*figure 2*). The second shoot growth period began during the second rapid

fruit growth period. It is clear that both peaks of shoot growth occurred simultaneously with the lowest rate of fruit growth.

The trunk grew from July to October after shoot and fruit growth was completed and coinciding with minimum root growth (data not shown). The trunk growth rate from January to July was constant at about  $1.1 \text{ mm month}^{-1}$ , whereas it was about  $2.6 \text{ mm month}^{-1}$  from July to October.

This pattern indicates that maximum growth periods of the three organs are distinct and take place sequentially. First of all, roots start to grow to absorb water and nutrients. This mobilizes reserves for the flowering process. After that, shoots began to grow, followed by maximum root growth and coinciding with the first fruit growth stage and lower values of shoot growth. This is followed by a second rapid fruit growth stage and finally by a secondary shoot growth period, which maximum occurred after harvest (*figure 2*).

According to *figure 3*, the abscission of buds (stage A), and reproductive organs (from B to F) occurred from the beginning of February to the end of April, with maximum values during March. Fruits started to drop in March (after fruit set), with a maximum at the beginning of April and a lower peak in May, just before harvest. Leaf abscission started at the beginning of November, with total leaf fall occurring at the end of the year.

#### **4. DISCUSSION**

The use of GDH accumulation for each phenological stage (Table II) enables comparisons to be made across different years and geographical areas, so that the beginning and duration of a given stage can be predicted on the basis of a

quantitative parameter, instead of a fixed calendar date [31, 37]. Apricot phenological stages were identified by a universal scale using a decimal code, the BBCH General Scale (*figure 1* and Table II), a nomenclature already used by several authors in different trees: pomegranate [33], citrus [3], quince [30], stone and pip fruits [32], as well as vegetables [27].

Although studies related to phenological stages in apricot trees are scarce, some authors, such as Burgos et al. [8] indicated that the stigma becomes receptive when it has accumulated 300 °C GDH, receptivity lasting 4-6 days. Maximum flowering for late apricot cultivars occurred at around 4900 °C GDH [29], whereas in Búlida (early-medium cultivar) maximum flowering occurred at around 3800 °C GDH (Table II).

Costes et al. [11] indicated that harvest in Rouge du Roussillon apricot started 110 days after full bloom. The use of number of days is an imprecise way of timing (the duration or the initiation) of a phenological process, since there may be substantial year-to-year variations in processes such as bloom or harvest, a problem that is avoided using energy units accumulation as the GDH model does [37]. In this sense Melgarejo et al. [33] indicated that the use of degree days allows for comparisons to be made between different years and geographical areas. Our results using GDH accumulation confirm this statement because only minor year-to-year variations were found (Table II). Standard deviations of the mean values are of the same order as those found for blooming time and related processes in peaches in different years [37].

It must be taken into account that, in spite of the relation between fruit maturity and GDH accumulation, fruit maturity is influenced by other factors,

such as cultivation techniques, water availability and fruit competition [35, 49].

Similar to the conclusions that can be drawn from *figure 3*, Tamássy and Zayan [45] indicated that there are two periods of abscission in the reproductive stages of apricot trees. The first stage was at bud swelling (the end of winter dormancy) and the other coincided with maximum flowering. The initial fruit abscission observed (*figure 3*) was due to an anomalous fruit set [14, 18]. The main period of fruit abscission, which coincides with the end of cell division (stage I of fruit growth), is due to competition for carbohydrate between fruits [13] and/or the lack of auxins [6]. Near-harvest fruits drop through the effect of fruit abscission layer formation [36].

Such an alternating root and shoot growth pattern (*figure 2*) has also been found in other fruit and forest trees [16, 23, 48]. In this sense, Ross and Catlin [38] indicated that the main almond root growth period occurs immediately after the shoot growth period. Our results in apricot trees indicate the existence of a first root growth period before bud burst and a second period between the first and the second shoot growth periods; in any case, the peaks of maximum growth of shoots and roots did not coincide (*figure 2*). Bevington and Castle [7] indicated that maximum orange shoot growth coincides with minimum root growth, due to the inhibitory effect of the auxins produced during active shoot growth [4].

A relative separation between the most intense periods of shoot and fruit growth (*figure 2*) has also be observed in pear and peach [10], pistachio [19], lemon [12] and mandarin trees [22]. Torrecillas et al. [46] pointed to a clear distinction between the main periods of shoot and fruit growth in apricot trees. This can have a clear advantage when irrigation has to be managed under

restricted water supply conditions, as is the case of regulated deficit irrigation strategies, since water deficit will affect only one of these processes at a time [20, 41]. In this sense, it has been reported that regulated deficit irrigation may be applied to control vegetative growth without having a negative impact on yield or crop revenue [9, 12, 21].

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### **REFERENCES**

- [1] Abrisqueta J.M., Hernansáez A., Franco J.A., Root dynamics of young almond trees under different drip-irrigation rates. *J. Hortic. Sci.* 69 (1994) 237-242.
- [2] Abrisqueta J.M., Ruiz A., Franco J.A., Water balance of apricot trees (*Prunus armeniaca* L., cv. Búlida) under drip irrigation. *Agric. Water Manag.* 50 (2001) 211-227.
- [3] Agustí M., Zaragoza S., Bleiholder H., Buhr L., Hack H., Klose R., Stauss R., Escala BBCH para la descripción de los estadios fenológicos del desarrollo de los agrios (Gén. Citrus). *Levante Agrícola* 332 (1995) 189-199.
- [4] Atkinson D., The distribution and effectiveness of the roots of tree crops. *Hortic. Rev.* 2 (1980) 424-490.

- [5] Baggiolini M., Stades repères de l'abricotier. Revue Romande d'Agriculture, Viticulture et Arboriculture 8 (1952) 28-29.
- [6] Baldini E., Arboricultura General. Mundi-Prensa. Madrid, 1992.
- [7] Bevington K.B., Castle W.S., Annual root growth pattern of young citrus trees in relation to shoot growth, soil temperature, and soil water content. J. Am. Soc. Hortic. Sci. 110 (1985) 840-854.
- [8] Burgos L., Egea J., Dicenta F., Effective pollination period in apricot (*Prunus armeniaca* L.) varieties. Ann. Appl. Biol. 119 (1991) 533-539.
- [9] Chalmers D.J., Mitchell P.D., van Heek L., Control of peach tree growth and productivity by regulated water supply, tree density, and summer pruning. J. Amer. Soc. Hort. Sci. 106 (3) (1981) 307-312.
- [10] Chalmers D.J., A physiological examination of regulated deficit irrigation. New Zealand J. Agric. Sci. 4 (1989) 44-48.
- [11] Costes E., Audubert A., Jaffuel S., Jay M., Demene M.N., Lichou J., Chronologie du développement du fruit en relation avec la croissance végétative chez l'abricotier *Prunus armeniaca* L. cv. Rouge du Roussillon. Can. J. Bot. 73 (1995) 1548-1556.
- [12] Domingo R., Ruiz-Sánchez M.C., Sánchez-Blanco M.J., Torrecillas A., Water relations, growth and yield of Fino lemon trees under regulated deficit irrigation. Irrig. Sci. 16 (1996) 115-123.
- [13] Egea J., El ciclo anual en frutales de zona templada. Factores de fructificación. Fruticultura Profesional 73 (1995) 34-39.
- [14] Egea J., Garcia J.E., Egea L., Berenguer T., Self-incompatibility in apricot cultivars. Acta Hortic. 293 (1991) 285-293.

- [15] Erez A., Couvillon G.A., Characterization of the influence of moderate temperatures on rest completion in peach. *J. Am. Soc. Hortic. Sci.* 112 (1987) 677-680.
- [16] Faust M., *Physiology of temperate zone fruit trees.* John Wiley and Sons. New York, 1989.
- [17] García J.E., Egea J., Egea L., Berenguer T. The Floral biology of certain apricot cultivars in Murcia. *Adv. Hort. Sci.* 2 (1988) 84-87.
- [18] Gil-Albert F., *Tratado de arboricultura frutal. Vol I: Aspectos de la morfología y fisiología del árbol frutal.* 3rd Ed. Mundi-Prensa. Madrid, 1989.
- [19] Goldhamer D.A., Phene B.C., Beede R., Scherlin L., Mahan S., Rose D., Effects of sustained deficit irrigation on pistachio tree performance. *California Pistachio Industry Annual Report 1986-87* (1987) 61-66.
- [20] Goldhamer D.A., *Drought Irrigation Strategies for Deciduous Orchards.* Cooperative Extensión. University of California, Division Agricultural and Natural Resources. Publication nº 21453, 1989.
- [21] Goldhamer D.A., Regulated deficit irrigation of fruit and nut trees. *International Water & Irrigation Review* 17 (4) (1997) 14-19.
- [22] González-Altozano P., Castel J.R., Regulated deficit irrigation in 'Clementina de Nules' citrus trees. II. Vegetative growth. *J. Hortic. Sci. Biotech.* 74 (2000) 706-713.
- [23] Harris J.R., Bassuk N.L., Zobel R.W., Whitlow T.H., Root and shoot growth periodicity of green ash, scarlett oak, Turkish hazelnut and tree lilac. *J. Am. Soc. Hortic. Sci.* 120 (1995) 211-216.



- [24] Huguet J.G., Li S.H., Defrance H., Influence de la disponibilité en eau du sol sur la qualité des fruits chez le pêcher *Prunus persica* L. 9<sup>o</sup> Colloque sur les *Recherches Fruitières*. Avignon. (1990) 135-144.
- [25] Irving D.J., Drost J.H., Effects of water deficit on vegetative growth, fruit growth and fruit quality in Cox's orange Pippin apple. *J. Hort. Sci.* 62 (1987) 427-432.
- [26] Lampinen B.D., Shackel K.A., Southwick S.M., Olson B., Yeager J.T., Goldhamer D.A. Sensitivity of yield and fruit quality of french prune to water deprivation at different fruit growth stages. *J. Amer. Soc. Hort. Sci.* 120 (1995) 139-147.
- [27] Lancashire P.D., Bleiholder H., van den Boom T., Langelüddeke P., Stauss R., Weber E., Witzemberger A., A uniform decimal code for growth stages of crops and weeds. *Ann. Appl. Biol.* 119 (1991) 561-601.
- [28] Larson K.D., Dejong T.M., Johnson R.S., Physiological and growth responses of mature peach trees to postharvest water stress. *J. Amer. Soc. Hort. Sci.* 113 (1988) 296-300
- [29] Legave J.M., García, G., Marco F., Interference of temperature conditions and the varietal requirements in cold and heat, on the determinations of the end of dormancy, then the blooming, of diverse varieties of apricot trees in a French cultivation area. *Fruits* 39 (6) (1984) 399-410.
- [30] Martínez-Valero R., Melgarejo P., Salazar D.M., Martínez R., Martínez J.J., Fernández F., Phenological stages of the quince tree (*Cydonia oblonga*). *Ann. Appl. Biol.* 139 (2001) 189-192.

- [31] McIntyre G.N., Lider L.A., Ferrari N.L., The chronological classification of grapevine phenology. *Am. J. Enol. Vitic.* 33 (1982) 80-85.
- [32] Meier U., Graf H., Hack H., Hess M., Kennel W., Klose R., Mappes D., Seipp D., Stauss R., Streif J., van den Boom T., Phänologische entwicklungsstadien der kernobsten (*Malus domestica* Borkh. und *Pyrus communis* L.), des steinobstes (*Prunus-Arten*) der johannisbeere (Ribes-Arten) und der erdbeere (*Fragaria x ananassa* Duch.). Codierung und beschreibung nach der erweiterten BBCH-Skala, mit abbildungen. *Nachrichtenblatt Deutsche Phanzenschutzd* 46 (1994) 141-153.
- [33] Melgarejo P., Martínez-Valero R., Guillamón J.M., Miró M., Amorós A., Phenological stages of the pomegranate tree (*Punica granatum* L.). *Ann. Appl. Biol.* 130 (1997) 135-140.
- [34] Mitchell P.D., Chalmers D.J., The effect of reduced water supply on peach tree growth and yields. *J. Amer. Soc. Hort. Sci.* 107 (1982) 853-856.
- [35] Morris J.R., Cawthon D.L., Effects of irrigation, fruit load and potassium fertilization on yield, quality and petiole analysis on Concord grapes. *Am. J. Enol. Vitic.* 33 (1982) 145-148.
- [36] Pérez-Pastor A., Estudio agronómico y fisiológico del albaricoquero en condiciones de infradotación hídrica. Tesis Doctoral. Universidad Politécnica de Cartagena, Murcia, 2001. 235 pp.
- [37] Richardson E.A., Seeley S.D., Walker R.D., Anderson J., Ashcroft G., Pheno-climatography of spring peach bud development. *HortScience* 10 (1975) 236-237.

- [38] Ross N.W., Catlin P.B., Rootstocks and root physiology, in: Almond Orchard Management. University of California. Division Agricultural Science, 1978, pp. 25-29.
- [39] Ruggiero C., Consumo idrico dell'albicocco irrigato a goccia, per aspersione e non irrigato durante i primi cinque anni dall'impianto. Riv. Ortoflorofruttic. It. 70 (1986) 1-11.
- [40] Ruiz-Sánchez M.C., Egea J., Galego R., Torrecillas A., Floral biology of Búlida apricot trees subjected to postharvest drought stress. Ann. Appl. Biol. 135 (1999) 523-528.
- [41] Sánchez-Blanco M.J., Torrecillas A., Aspectos relacionados con la utilización de estrategias de riego deficitario controlado en cultivos leñosos, in: Riego Deficitario Controlado. Fundamentos y Aplicaciones. Colección Cuadernos VALUE 1. Mundi Prensa/Unión Europea, 1995, pp. 43-63.
- [42] Sánchez-Capuchino J.A., Contribución al conocimiento de necesidades en frío invernal de variedades frutícolas. Levante Agrícola 61 (1967) 26-28.
- [43] Shalhevet J., Mantell A., Bielorai H., Shimshi D. Irrigation of Field and Orchard Crops under Semi-arid Conditions. *IIIC. n° 1 (revised versión)*. Israel. Canada. (1979) 124 pp.
- [44] Tabuenca M.C., Herrero J., Influencia de la temperatura en la época de floración de los frutales. An. Aula Dei 8 (1966) 115-153.
- [45] Tamássy I., Zayan M., Critical temperatures in winter (after rest period) and in spring (at blooming time) for fruit buds and open flowers of some apricot varieties from different groups. Acta Hortic. 121 (1982) 63-67.

- [46] Torrecillas A., Domingo R., Galego R., Ruiz-Sánchez M.C., Apricot tree response to withholding irrigation at different phenological periods. *Sci. Hortic.* 85 (2000) 201-215.
- [47] Upchurch D.R., Ritchie J.T., Root observations using a video recording system in mini-rhizotrons. *Agron. J.* 75 (1983) 1009-1015.
- [48] Watson G.W., The relationship of root growth and tree vigour following transplanting. *Arboric. J.* 11 (1987) 97-104.
- [49] Williams L.E., Grimes D.W., Modelling vine growth-development of a data set for a water balance subroutine. *Proceeding of the 6th Australian Wine Industry Technology Conference Adelaide, 1986.* Australian Industrial Publisher, 1987, pp. 169-174.

## FIGURE LEGENDS

**Photograph 1.** Phenological growth stages of Búlida apricot trees, according to BBCH (numbers) and Baggiolini (letters) codes.

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**Figure 1.** Reproductive stages according to Baggiolini code (flower buds) of Búlida apricot trees under non-limiting conditions. 1997/98. Each box plot is the average of eight replicates. Vertical line in the box line indicates the median. Points outside the box are  $\pm$  Std.

**Figure 2.** Appearance of new roots, expressed as root length density (RLD, cm cm<sup>-3</sup> soil), shoot growth rate (SGR, cm day<sup>-1</sup>) and fruit growth rate (FGR, g day<sup>-1</sup>) in Búlida apricot trees under non-limiting conditions. Full bloom 1997: 5 March, 1998: 8 March. Harvest: mid may. Each point is the average of eight replicates  $\pm$  Std

**Figure 3.** Fall of buds (stage A), reproductive organs (stages B to F), fruits and leaves of Búlida apricot trees under non-limiting conditions. 1997/98. Each point is the average of eight replicates of different elements collected in the catching nets, on a dry weight basis  $\pm$  Std.

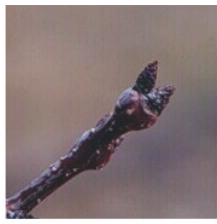
Table I. Monthly maximum, minimum and mean air temperatures, relative humidity (RH), rainfall, mean daily evaporation rate (Epan), wind speed (2 m) and net radiation.

Month	Temperature (°C)			RH (%)	Rainfall		Epan	Wind	Net radiation
	Max.	Min.	Mean	Mean	mm	days	mm d <sup>-1</sup>	km d <sup>-1</sup>	W m <sup>-2</sup>
<b>1997</b>									
Jan	13.1	7.6	10.4	90.5	42.3	9	1.29	105.4	74.6
Feb	18.4	8.9	13.6	78.9	1.3	1	2.87	102.5	150.9
Mar	20.5	8.6	14.5	74.9	24.0	1	3.72	82.6	205.6
Apr	20.6	10.5	15.5	85.2	92.3	6	3.10	79.5	209.4
May	24.3	13.1	18.7	78.2	13.6	1	5.22	91.5	270.5
Jun	28.2	16.5	22.3	74.3	65.8	3	6.75	85.1	273.4
Jul	29.7	16.9	23.2	74.0	0	0	7.22	87.0	287.2
Aug	29.4	18.2	23.8	76.2	0	0	6.14	81.0	250.9
Sep	26.9	15.3	21.1	71.7	135.8	5	3.87	144.0	208.3
Oct	23.8	13.9	19.2	86.8	12.3	2	2.86	47.8	147.5
Nov	18.4	9.6	14.0	84.7	20.7	5	1.77	61.8	99.1
Dec	15.7	8.1	11.8	84.2	28.6	5	1.23	94.3	72.4
<b>1998</b>									
Jan	14.6	6.9	10.8	85.3	10.5	1	1.87	145.2	82.5
Feb	14.9	7.8	11.3	88.9	32.5	2	1.82	162.1	137.9
Mar	20.4	9.1	14.7	71.6	4.0	1	3.85	106.2	186.5
Apr	21.3	9.6	15.4	65.9	0	0	4.70	96.3	219.1
May	22.2	12.2	17.2	83.0	45.0	2	4.61	99.2	275.3
Jun	29.1	16.1	22.6	76.3	0	0	6.24	103.7	291.3
Jul	32.4	18.1	25.3	72.2	0	0	7.70	112.4	300.6
Aug	30.3	18.3	24.3	85.5	0	0	6.76	108.7	262.8
Sep	27.9	17.9	22.9	63.3	23.0	1	4.66	90.4	206.3
Oct	22.3	12.4	17.3	58.9	0	0	2.95	79.5	139.3
Nov	19.2	5.6	12.4	68.5	34.5	1	1.96	109.1	102.9
Dec	15.5	4.4	9.9	71.6	53.0	1	1.54	91.8	69.2

Table II. Pheno-climatology for Búlida apricot. Predominant dates of growth stages, occurrence of BBCH codes, and growing degree hour (GDH) accumulated at the beginning of each stage.

<b>BBCH code</b>	<b>Growth stage</b>	<b>Date</b>	<b>Duration days</b>	<b>GDH (°C) Initial*</b>
00	Bud completely closed covered by brown scales	< 15 Feb	-	-
51	Flower bud swelling. Buds are closed and light scales are perceptible.	15-25 Feb	11	0
01	Beginning of vegetative bud swelling	21-25 Feb	5	1035 ± 58
09	Bud open and green leaf tip perceptible	26-27 Feb	2	1827 ± 77
55	Calyx perceptible, sepals closed forming a red ball	26 Feb-4 Mar	7	1827 ± 77
10	First leaves emerging	28 Feb-5 Mar	6	2153 ± 213
31	Beginning of the first shoot growth, internodes lengthen.	1-12 Mar	12	2334 ± 262
58	Flower petals perceptible, forming a hollow ball	5-6 Mar	2	3274 ± 319
11-15	Expanding leaves, not yet full size	6-12 Mar	7	3504 ± 275
59	Flower starting to open and the pistil anther perceptible	7 Mar	1	3784 ± 283
65	Full bloom, 50 % of open flowers	7-9 Mar	3	3784 ± 283
67	Flower fading	8-14 Mar	7	4125 ± 402
69	End of flowering, all petals fallen	10-14 Mar	5	4725 ± 413
71	Fruit set, beginning of ovary growth	10 Mar-1 Apr	23	4725 ± 413
19	Fully expanded leaves	13-20 Mar	8	5150 ± 337
32-37	Shoots about 20 and 70 % of the final length	20 Mar-2 Apr	14	6596 ± 357
72	Green fruits surrounding by a wilted sepal crown	24 Mar-8 Apr	16	7177 ± 338
73	Physiological fruit drop	2-9 Apr	8	9269 ± 329
75	Fruits about 50 % of final size	10-14 Apr	5	11234 ± 735
79	Fruits about 90 % of final size	15-27 Apr	13	11886 ± 386
81	Beginning of fruit colouring	28 Apr-8 May	11	15124 ± 447
89	Fruit ripe for consumption, and showing full organoleptic characteristics	26 May-3 Jun	9	23116 ± 573
39	Second shoot growth	20 May-21 Jul	63	21332 ± 686
93	Beginning of senescence	10-20 Nov	11	86139 ± 1831
95	Leaves turned yellowish and started to fall	20-30 Nov	11	88377 ± 1976
97	Total leaf abscission. Winter dormancy period	15 Dec	-	93367 ± 1901

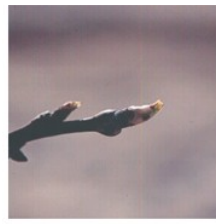
\*Growing degree hour (GDH) accumulation after completion of rest as determined from time of 600 chill unit accumulation. Values are mean of the two years ± Std



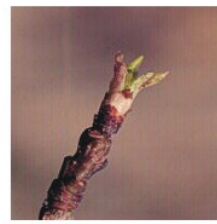
**A**  
00



01



07



09



10



11



12



15



19



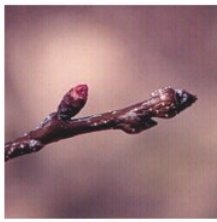
31



32



35



**B**  
51



**C**  
55



**D**  
58



**E**  
59



61



**F**  
65



**G**  
67



69



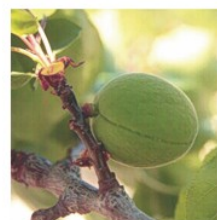
**H**  
71



**I**  
72



73



75



89



93



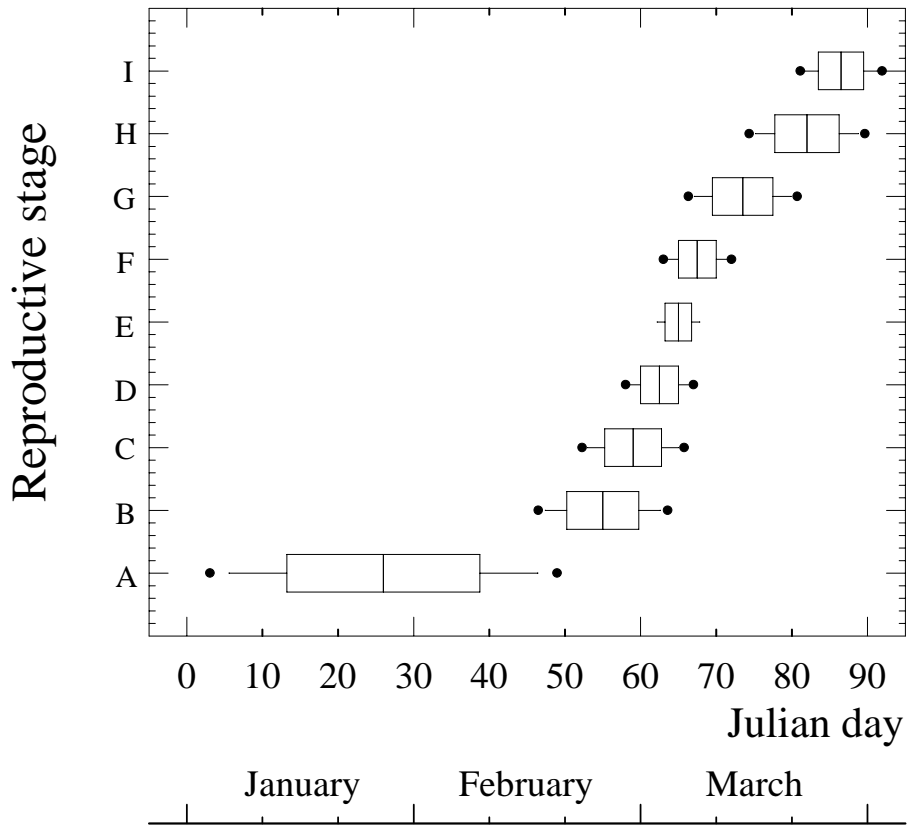
95



97

Photograph 1.





**Figure 1.**

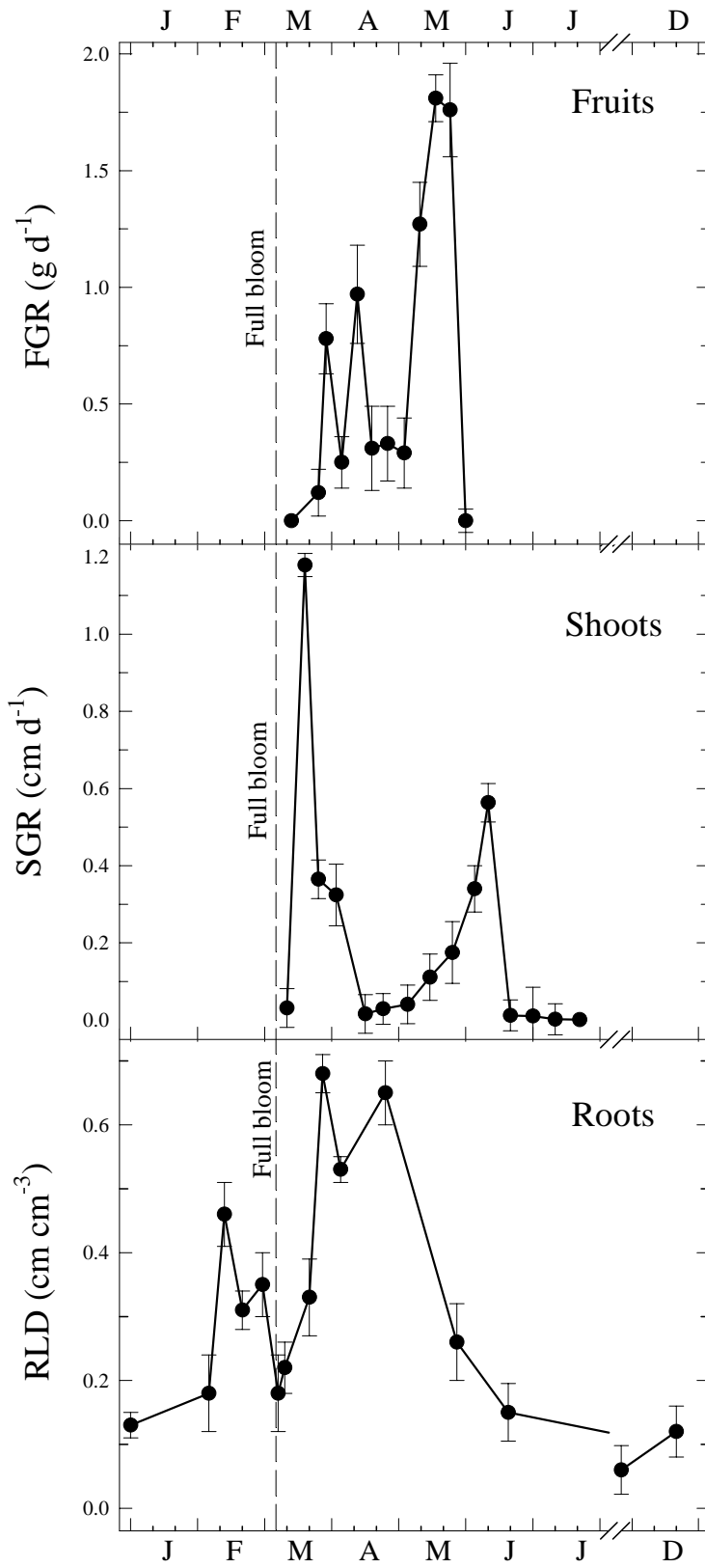


Figure 2.

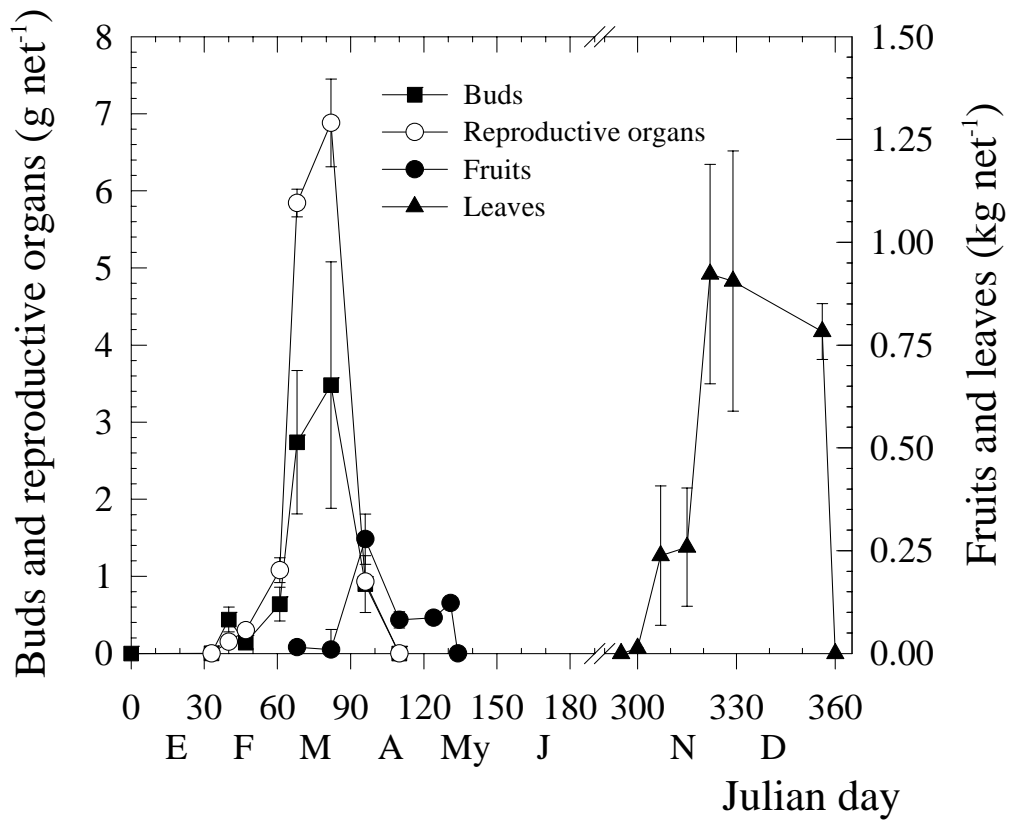


Figure 3.